

# A neuromorphic model of active vision shows spatio-temporal encoding in lobula neurons can aid pattern recognition in bees

HaDi MaBouDi<sup>1,2,3\*</sup>▲, Mark Roper<sup>3,4</sup>, Marie-Genevieve Guiraud<sup>3,5</sup>, Lars Chittka<sup>3</sup>, James A.R. Marshall<sup>1,2</sup>▲

<sup>1</sup>Department of Computer Science, University of Sheffield, Sheffield, UK

<sup>2</sup>Neuroscience Institute, University of Sheffield, Sheffield, UK

<sup>3</sup>School of Biological and Chemical Sciences, Queen Mary University of London, London, UK

<sup>4</sup>Drone Development Lab, Ben Thorns Ltd, Colchester, Essex, UK

<sup>5</sup>School of Natural Sciences, Macquarie University, North Ryde, NSW, Australia

\*Corresponding author: HaDi MaBouDi, [maboudi@gmail.com](mailto:maboudi@gmail.com)

▲ Current address: Opteran Technologies, Sheffield Innovation Centre, Sheffield, UK

**Short title:** Insect-inspired minimal model for active vision

## Abstract

Bees possess remarkable cognitive abilities in on-the-fly visual learning, making them an ideal model for studying active information acquisition and representation. In this study, we investigated the minimal circuitry required for active vision in bees by considering their flight behaviours during visual pattern scanning. By developing a neural network model inspired by the insect visual system, we examined the influence of scanning behaviour on optic lobe connectivity and neural activity. Through the incorporation of non-associative learning and exposure to diverse natural images, we obtained compelling results that align with neurobiological observations. Our findings reveal that active scanning and non-associative learning dynamically shape the connectivity within the visual lobe, resulting in an efficient representation of visual input. Interestingly, we observed self-organization in orientation-selective neurons in the lobula region, characterized by sparse responses to orthogonal bar movements. These dynamic orientation-selective cells cover various orientations, exhibiting a bias towards the speed and contrast of input sampling. To assess the effectiveness of this spatiotemporal coding for pattern recognition, we integrated our model with the mushroom body circuitry underlying associative learning. Notably, our model demonstrated impressive performance across several pattern recognition tasks, suggesting a similar coding system within the bee visual system. Overall, this study integrates behavioural experiments, neurobiological findings, and computational models to reveal how complex visual features can be condensed through spatiotemporal encoding in the lobula neurons, facilitating efficient sampling of visual cues for identifying rewarding foraging resources. Our findings have broader implications for understanding active vision in diverse animals, including humans, and offer valuable insights for the application of bio-inspired principles in the design of autonomous robots.

## Keywords

active vision, image statistics, lobula, mushroom bodies, non-associative learning, orientation selective neurons, scanning behaviour, visual recognition.

## 38 Introduction

39 Bees are capable of remarkable cognitive feats, in particularly in visual learning (Srinivasan, 2010; Turner,  
40 1911; Von Frisch, 1914; Wehner, 1967); they can not only learn to associate a colour or orientation of a bar  
41 with reward (Dyer et al., 2011; Srinivasan, 1994; Stach et al., 2004) but are also able to identify specific features  
42 to categorise visual patterns, by finding the relevant stimuli properties (Benard et al., 2006; Stach et al., 2004).  
43 Furthermore, bees have demonstrated the capacity to grasp abstract concepts (Avarguès-Weber et al., 2011;  
44 Giurfa et al., 2001; Guiraud et al., 2018; MaBouDi et al., 2020c; Menzel, 2012) and solve numerosity tasks by  
45 scanning the elements within the presented stimuli (MaBouDi et al., 2020a). These exceptional capabilities  
46 position bees as a valuable animal model for investigating the principles of visual learning through the analysis  
47 of their behavioural responses (Menzel and Giurfa, 2006; Srinivasan, 2010). Nevertheless, it is still unclear how  
48 bees, with low visual acuity (Gribakin, 1975; Srinivasan and Lehrer, 1988) and limited neural resources can  
49 recognise complex patterns, and indeed perceive the visually intricate natural world they encounter during  
50 daily foraging activities (Chittka and Niven, 2009; Giurfa, 2013).

51

52 Bees, as vital pollinators, must cope with the variability of the natural environment for their survival. The  
53 natural scenes that animals typically encounter are structured differently to random/non-natural scenes  
54 (Matthews et al., 2018; Ruderman, 1994; Simoncelli and Olshausen, 2001; Zimmermann et al., 2018). It has  
55 been hypothesised that visual sensory neurons must efficiently adapt to the regularities of the natural scene  
56 to take advantage of this spatio-temporal structure, and to efficiently code the information in the visual  
57 environment (Barlow, 1961). Hence, Insects visual neurons have evolved to provide robust and efficient  
58 responses to naturalistic inputs, enabling survival in complex ecological niches (Dyakova et al., 2019, 2015;  
59 Dyakova and Nordström, 2017). Numerous studies have highlighted the remarkable adaptability of the sensory  
60 pathway in insects, demonstrating robust response and behaviour across various input parameters, such as  
61 contrast, spatial frequency and spatiotemporal correlations (Arenz et al., 2017; Brinkworth and O'Carroll,  
62 2009; Clark et al., 2014; Dyakova et al., 2019; Dyakova and Nordström, 2017; Juusola and Song, 2017;  
63 Schwegmann et al., 2014; Serbe et al., 2016; Song and Juusola, 2014, 2014; Van Hateren, 1997; van Hateren,  
64 1992). For instance, Song and Juusola (2014) showed that fly photoreceptors extract more information from  
65 naturalistic time series compared to artificial stimuli or white noise, exhibiting stronger responses with a higher  
66 signal-to-noise ratio (Song and Juusola, 2014). Despite insights gained from the statistical properties of the  
67 natural scene, the precise neural mechanisms underlying visual processing of natural scene remain elusive,  
68 necessitating further investigation. This study aims to investigate the theoretical aspects of how insect visual  
69 circuitry adapts to regularities in natural scene. One of our objectives is to understand the efficient coding  
70 strategies and robust response mechanisms employed by these neurons that play a crucial role in enhancing  
71 visual pattern recognition and facilitate survival and navigation in variable environments.

72 In the realm of animal vision, active vision strategies have been observed, where animals actively scan targets  
73 and extract visual information over time (Land, 1999; Land and Nilsson, 2012; Yarbus, 2013). Primates employ  
74 eye movements, including microsaccades, as an effective sampling strategy that enhances fine spatial  
75 information and improves the encoding details of natural stimuli (Anderson et al., 2020; Land, 1999; Rucci et  
76 al., 2007; Rucci and Victor, 2015). Other species, such as insects, exhibit active vision strategies through  
77 characteristic head/body-movements or a certain approaching paths during a visual tasks (Chittka and  
78 Skorupski, 2017; Dawkins and Woodington, 2000; Land, 1973; Land and Nilsson, 2012; Langridge et al., 2021).  
79 Recent studies have discovered that *Drosophila* are able to move their retinas to stabilise their retinal images,  
80 achieving hyperacute vision and enhancing depth perception (Fenk et al., 2022; Juusola et al., 2017).  
81 Honeybees may need to sample and integrate colour information due to their limited ability to discriminate  
82 similar colours in brief flashes (<50ms) (Nityananda et al., 2014). They exhibit a sequence of movements in  
83 response to particular visual stimuli and employ continuous sampling to build a representation of their  
84 environment (Boeddeker et al., 2015; Collett et al., 1993; Doussot et al., 2021; Guiraud et al., 2018; Langridge  
85 et al., 2018; Lehrer and Collett, 1994; MaBouDi et al., 2020a; Werner et al., 2016). For instance, bumblebees  
86 acquire visual details of numerosity by sequential scanning of stimulus elements rather than parallel  
87 processing to enumerate the countable elements within the visual displays (MaBouDi et al., 2020a). A recent  
88 analysis of bumblebees' flight trajectories showed that they sequentially scanned specific regions of the  
89 patterns prior to making a decision, instead of global pattern processing (Langridge et al., 2021; MaBouDi et  
90 al., 2021b). Therefore, the low-resolution compound eyes and the possibly reduced parallel processing in  
91 insects (compared to vertebrates) suggest that these bees may employ an active vision strategy by continuous  
92 sampling to build up a picture of their environment (Chittka and Skorupski, 2017; Nityananda et al., 2014).  
93 These active vision strategies, akin to primate eye movements, serve fundamental functions in early visual  
94 processing for redundancy reduction (Doussot et al., 2021; Kuang et al., 2012; Odenthal et al., 2021).  
95 Nonetheless, it is still poorly understood how active vision strategies enable bees to overcome their limited  
96 representational capacity, discover regularities in the visual input and solve complex visual discriminations.

97 Taking advantage of the bee's visual ability and the detailed information on flight's paths analysed in our  
98 previous study when bees solved a simple visual task (MaBouDi et al., 2021b), we conducted further research  
99 to investigate the necessary and minimally sufficient circuitry required for active vision of achromatic pattern  
100 recognition in bees. Importantly, our primary objective was to investigate how the scanning behaviour of bees  
101 contributes to the functional organization and connectivity of neurons in the visual lobe. We specifically  
102 focused on the hypothesis that complex visual features can be efficiently encoded through spatio-temporal  
103 patterns of activity in the lobula neurons, leading to distinct and specific representations necessary for learning  
104 in the miniature brain of bees. Through our study, we aimed to provide valuable insights into the intricate  
105 interplay between scanning behaviour and neural activity within the bee visual system, ultimately advancing

106 our understanding of the mechanisms underlying active vision. To achieve this, we developed a neuromorphic  
107 model of the bee optic lobes, incorporating the concept of efficient coding through the implementation of  
108 non-associative plasticity. Through this model, we demonstrated how spatial scanning behaviour of bees in  
109 response to naturalistic visual inputs shape the connectivity within the medulla (2<sup>nd</sup> optic ganglion) and  
110 facilitate an efficient representation of these inputs in the lobula (3<sup>rd</sup> optic ganglion). This efficiency is achieved  
111 through the self-organization of a specific set of orientation-selective neuron in the lobula, highlighting the  
112 combined impact of scanning behaviour and non-associative learning on shaping the neural circuitry within  
113 the bees' optic lobes. To assess the model, we integrated the optic lobe model with a secondary decision-  
114 making neural network inspired by neural mechanisms of associative learning in insect brains and supported  
115 by previous neurobiological findings (Cassenaer and Laurent, 2012; Okada et al., 2007; Paulk et al., 2009; Paulk  
116 and Gronenberg, 2008; Yang and Maddess, 1997). Visual input flight dynamics for the model were derived  
117 from our observations of bee behaviour during a visual discrimination task (MaBouDi et al., 2021b). This  
118 allowed us to evaluate and test the hypothesis of active sampling from our model against real-world behaviour  
119 results (MaBouDi et al., 2021b), as well as other published visual discrimination tasks performed by bees  
120 (Benard et al., 2006; Dyer et al., 2005; Srinivasan, 2010, 1994; Zhang and Horridge, 1992). Furthermore, we  
121 conducted a detailed analysis comparing the features and properties of neural responses that emerged in our  
122 model with existing neurobiological findings (James and Osorio, 1996; Paulk et al., 2008; Seelig and Jayaraman,  
123 2013; Yang and Maddess, 1997). By aligning our model's outputs with established neurobiological data, we  
124 enhanced the credibility and reliability of our model in accurately capturing essential aspects of neural  
125 processing associated with active vision.

126

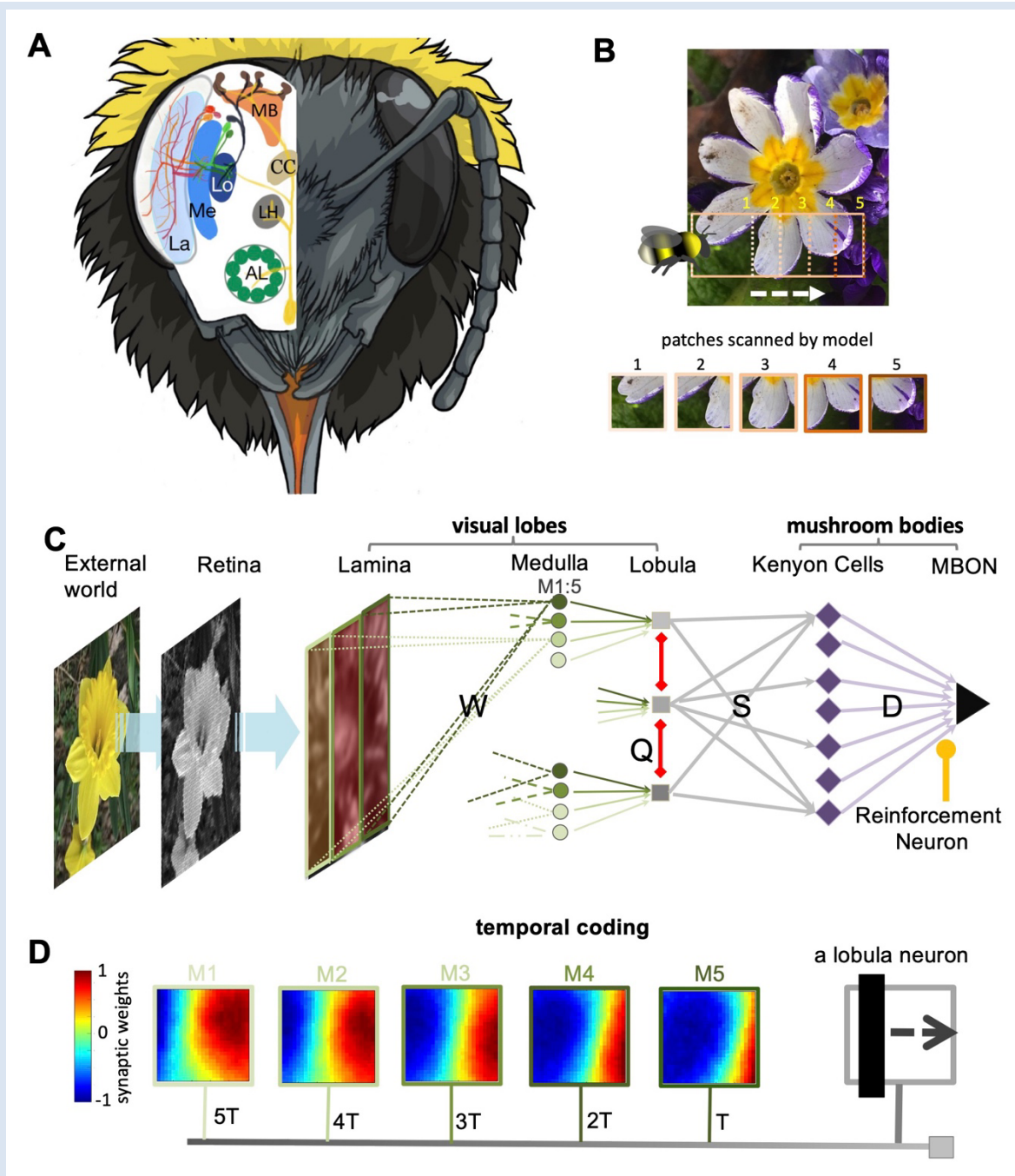
## 127 **Results**

### 128 ***A bio-inspired neural network of active vision***

129 To understand how bee scanning behaviour may efficiently shape the activity of neurons in the visual lobes of  
130 the bee brain based on the efficient coding, and how the visual information is processed for pattern  
131 recognition, we designed a neural network that drew inspiration from the known morphological and functional  
132 features of the insect brain (Figure 1A, C). The network approximates the neural circuitry which initially  
133 processes the visual input within the bees' lamina and medulla (1st and 2nd optic ganglia). Then, to replicate  
134 temporal encoding during scans (Figure 1B), we supposed a structure of time delay of between 1 - 5 'temporal  
135 instances' from the medulla neuron outputs to the lobula (3rd optic ganglion) wide-field neurons (Figure 1D).  
136 This arrangement enabled sequential sampling of specific locations along the scan line of the viewed visual



137 pattern, resulting in a gradually accumulated internal representation of the visual input as the ultimate output  
 138 of the lobula neurons.  
 139



**Figure 1. Neural network of active vision inspired by neurobiology and flight dynamics of bees. (A)** The right side displays the front view of the bumblebee head showing the component eye and antenna. Left hand side presents a schematic view of the bee's brain regions. Part of neural pathways from the retina to the mushroom bodies are also represented. Labels: AL – Antennal lobe; LH – Lateral horn; CC – Central complex; La – Lamina; Me – Medulla; Lo – Lobula; MB – Mushroom body. Figure was designed by Alice Bridges **(B)** A representation of the modelled bee's scanning behaviour of a flower demonstrating how a sequence of patches project to the simulated bee's eye with lateral movement from left to right. Below

shows the five image patches sampled by the simulated bee. **(C)** Representation of the neural network model of active vision inspired by micromorphology of the bee brain that underlie learning, memory and experience-dependent control of behaviour. The photoreceptors located in the eye component are excited by the input pattern. The activities of photoreceptors change the membrane potential of a neuron in the next layer, Lamina. The lamina neurons send signals (through  $W$  connectivity matrix) to the medulla neurons to generate spikes in this layer. Each wide-field lobula neuron integrates the synaptic output of five small-field medulla neurons. The lobula neurons are laterally inhibited by local lobula interconnections (via  $Q$  connectivity matrix). Lobula neurons send axons into the mushroom body for connection with Kenyon Cells (KCs) through a random matrix of connectivity,  $S$ . The KCs all connect to a single mushroom body output neuron (MBON) through random synaptic connections  $D$ . A single reinforcement neuron (yellow neuron) modulates the synaptic weights between KCs and MBON by simulating the release of octopamine or dopamine when presented with specific visual stimuli (see Method section). **(D)** A temporal coding model that is proposed as the connectivity between medulla and lobula neurons. Each matrix shows the inhibitory (blue) and excitatory (red) connectivity between lamina neurons to a medulla neuron at a given time delay. In this model, the five small-field medulla neurons that are activated by the locally visual input, at different times of scanning, send their activities to a wide-field lobula neuron with a synaptic delay such that the lobula neuron receives all medulla input signals at the same instance (*i.e.* in the presented simulation the lobula neuron is maximally activated by the black vertical bar passing across the visual field from the left to right. Each underlying medulla neuron encodes the vertical bar in a different location of the visual field).

140

141 In more detail, the model sampled the image input into five sequential patches of 75x75 pixels with a speed  
142 of 0.1 m/s, equating to a lateral movement of 15 pixels between patches (an example of sequential scanning  
143 is shown in Figure 1B, see Method Section) (MaBouDi et al., 2021b). The green pixel intensities of each image  
144 patch modified the membrane potential of 75x75=5625 photoreceptors within the single eye of the simulated  
145 bee. These photoreceptor responses converged to 625 lamina neurons *via* recurrent neural connectivity.  
146 Lamina neurons provided post-synaptic connections to 250 small-field medulla neurons *via* simple feed-  
147 forward connectivity (Figure 1C, see Method Section). The medulla response from each image patch (of five  
148 patches that cover the image through the movement) is computed using a spiking neural model. Their spiking  
149 activities are then integrated into the synapses of their corresponding lobula neuron, with a time delay  
150 designed to ensure that the lobula neuron receives all the underlying medulla input signals at the same time.  
151 It is important to note that the proposed spatio-temporal coding is a simplification, as within the bee brain,  
152 this process would be achieved through dendritic and synaptic latency, or through intermediate neuron  
153 transmission within the medulla that is influenced by the non-associative learning in the visual lobe (Figure  
154 1C, D). We hypothesised that connectivity in the medulla and lobula could be modified by exposing the visual  
155 lobe to a series of time-varying images while incorporating non-associative learning rules and efficient coding  
156 principle (see Method Section for details).

157

158 The neural representation of the visual inputs was subsequently transmitted and processed in the mushroom  
159 body, which serves as the visual learning centres of the bee brain (Ehmer and Gronenberg, 2002; Li et al., 2017;  
160 Paulk and Gronenberg, 2008) (Figure 1C). For simplicity, a single mushroom body output neuron (MBON) is  
161 exposed, where the firing rate of this neuron expresses the simulated bee's preference for any given visual  
162 input. By updating the synaptic weights within the mushroom body, we were able to train the neural network  
163 to associate visual patterns as either positive (resulting in low MBON firing rates) or negative (high MBON  
164 firing rates; see Discussion). After the visual network underwent acquisitive modification through non-  
165 associate learning and extensive exposure to natural images, the whole network, as a simulated bee, was  
166 trained and tested with a variety of pattern recognition tasks from the published literature (Benard et al.,  
167 2006; Dyer et al., 2005; Srinivasan, 2010, 1994; Zhang and Horridge, 1992), in particular with the “plus” and  
168 “multiplication sign” patterns used with the real bumblebees reported in MaBouDi et al., 2021 (Figure 2A).  
169 Finally, to assess the performance of the proposed active vision model in various pattern recognin tasks, we  
170 analysed the MBON as it is supposed to function as a pre-motor area. In this context, a lower response from  
171 the MBON to a specific pattern would indicate the bees' performance towards that particular pattern.  
172 Consequently, after multiple training trials, we presumed the MBON acts as a decision neuron, showing a  
173 lower response to the chosen pattern and higher response to the rejected pattern in the tests. Note that no  
174 reinforcement, or synaptic updates were implemented during the testing phase.

175

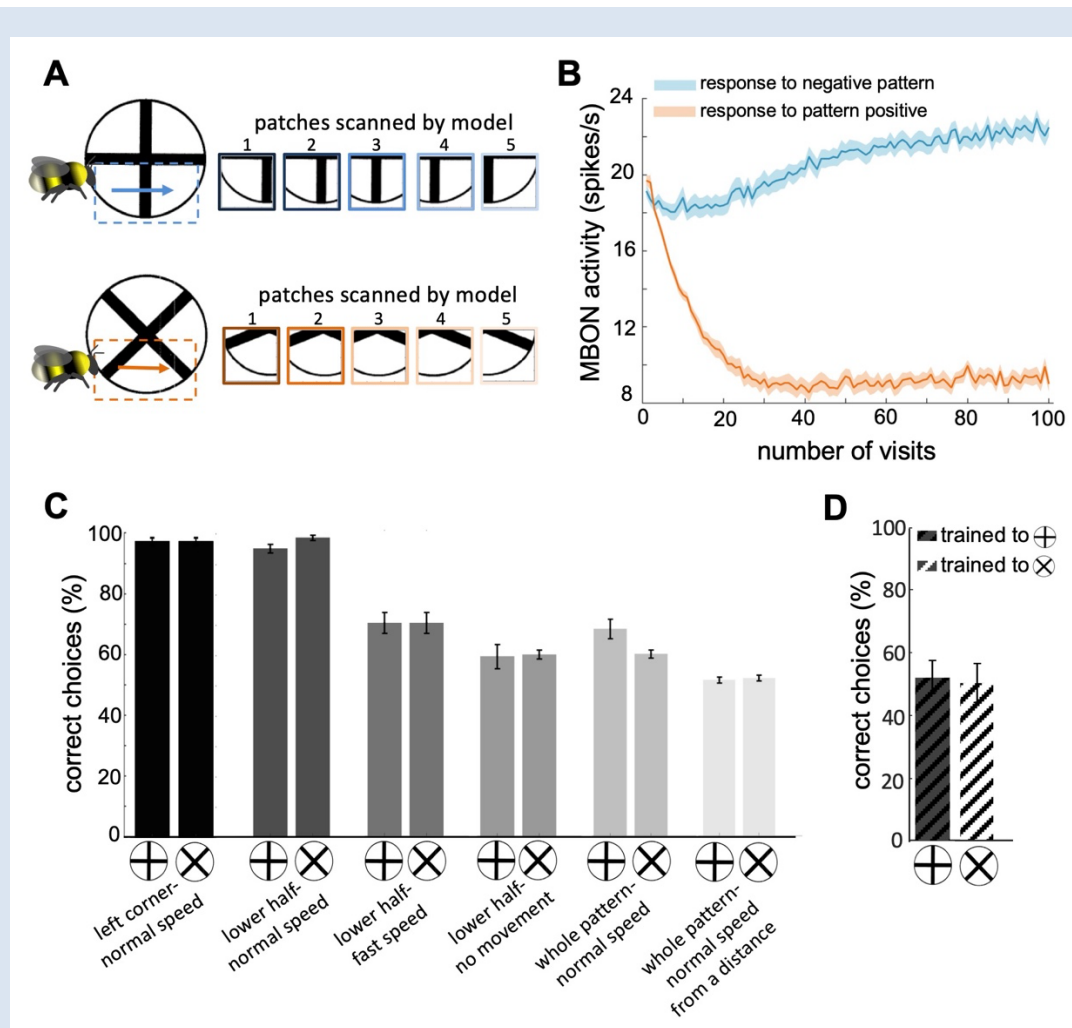
### 176 ***The model's performance in the same visual discrimination task is changed by scanning behaviour***

177 To replicate the bee behavioural results seen in the published literature (Benard et al., 2006; Dyer et al., 2005;  
178 MaBouDi et al., 2021b; Srinivasan, 1994, 2010; Zhang and Horridge, 1992), we implemented the type of  
179 computational plasticity in the mushroom body circuitry that is necessary to mediate both appetitive and  
180 aversive value encoding. Here, we assumed that classical spike-timing-dependent plasticity (STDP) (Figure 7)  
181 modulated by dopamine governs the plasticity rule between the mushroom body Kenyon cells (KC) and  
182 extrinsic Mushroom Body Output Neurons (MBON) in the presence of negative (or unrewarded) patterns.  
183 Additionally, we introduced a novel plasticity rule using STDP (Figure S2B) modulated by octopamine, which  
184 we hypothesised leads to synaptic depression among KC-MBON connections. These different plasticity rules  
185 were employed to investigate the synaptic dynamics in response to positive and negative patterns (see  
186 Method and Discussion sections).

187

188

189



**Figure 2. Simulated bees' performance in a pattern recognition task using different scanning strategies.**

Twenty simulated bees, with random initial neuronal connectivity in mushroom bodies (see Methods) and a fixed connectivity in the visual lobe that were shaped from the non-associative learning, were trained to discriminate plus from a multiplication symbol (100 random training exposures per pattern). The simulated bees scanned different regions of the patterns at different speeds. **(A)** Top and bottom panels show the five image patches sampled from the plus and multiplication symbols by simulated bees, respectively. It is assumed that the simulated bees scanned the lower half of the patterns with lateral movement from left to right with normal speed (0.1 m/s). **(B)** The plot shows the average responses of the MBON to rewarding multiplication and punishing plus pattern during training procedure (plus symbol rewarding, producing an Octopamine release by the reinforcement neuron, and the multiplication symbol inducing a Dopamine release). This shows how the response of the MBON to the rewarding plus was decreased while its response to the punishing multiplication pattern was increased during the training. The MBON equally responded to both multiplication and plus before the training (at number of visits =0). **(C)** The performance of the simulated bees in discriminating the right-angled plus and a 45° rotated version of the same cross (i.e. multiplication symbol) (MaBouDi et al., 2021b; Srinivasan, 1994), when the stimulated bees scanned different regions of the pattern (left corner, lower half, whole pattern) at different speeds: no speed 0.0m/s (i.e. all medulla to lobula temporal slices observed the same visual input), normal speed at 0.1 m/s and fast speed at 0.3m/s), and from a simulated distance of 2cm from stimuli (default) and 10cm (distal view). The optimal model parameters were for the stimulated bees at the default distance when only a local region of the pattern (bottom half or lower left quadrant) was scanned at a normal speed. **(D)** Mean performance ( $\pm$ SEM) of two groups of simulated bees in

discriminating the plus from multiplication patterns when their inhibitory connectivity between lobula neurons were not modified by non-associative learning rules.

190

**Video 1: An example of bee's flight in recognising a plus sign pattern.** The bee underwent training to receive 10  $\mu$ l 50% sucrose solution (w/w) from the feeding tubes at the centre of the plus pattern. After carefully inspecting the lower half of the plus sign, the bee accurately chose the correct pattern (MaBouDi et al., 2021b).

191

192

**Video 2: An example of bee's flight in recognising a multiplication sign pattern.** The bee underwent training to receive 10  $\mu$ l 50% sucrose solution (w/w) from the feeding tubes at the centre of the multiplication sign pattern. After carefully inspecting the lower half of the pattern, the bee accurately chose the correct pattern (MaBouDi et al., 2021b).

193

194 We first trained the model using a differential conditioning task in which the correct stimulus S+ is paired with  
195 reward while incorrect stimulus S- is delivered with punishment. Hereafter for simplicity, we use a subscript S  
196 to the label of valence of the pattern with + for positive and - for negative valence. In our simulations of  
197 associative learning, we update only those synaptic weights between KCs and MBON that correspond to the  
198 presented patterns (see Method section). To represent the learning task for different bees, we replicated the  
199 simulations, using different initial parameters (random neural connectivity between lobula and Kenyon cells  
200 and KCs-MBON connections). We then tested the performance of our model in a set of different visual task  
201 paradigms detailed below (Figures 2, 3). During the initial experiment where bees had to distinguish a plus  
202 from a multiplication sign, trained on the lower half of the plus (Figure 2A), the firing rate of the MBON  
203 decreases after presenting the plus (S+) and tends to increase after presenting S- (lower half of multiplication  
204 sign); whereas MBON responded equally to both plus and cross before training (Figure 2B). This indicates that  
205 the model can discriminate visual patterns S+ and S- using temporal coding and active scanning of patterns  
206 (Figure 2C). Conversely, the model with fixed random inhibitory connectivity in the lobula could not  
207 discriminate between the plus and the multiplication sign patterns (Figure 2D). This emphasizes the  
208 importance of the structured connectivity that emerges in the bee visual lobes by the non-associative learning  
209 (i.e. spatio-temporal receptive fields of lobula neurons), and subsequent performance of the simulated bees  
210 in visual learning tasks. In other words, rewarding patterns induce decreased extrinsic neuron responses, but  
211 result in higher responses for punished patterns (Figure 2B). This behaviour aligns with the biological  
212 observations in the alpha lobe of mushroom body PE1 neurons, which indicate a decrease in the response of  
213 MBON in the presence of the positive stimuli (Okada et al., 2007).

214

215 The replication of the plus and multiplication experiment with our simulated bees initially resulted in a poor  
216 performance compared to the real bees (Figure 2C), with the correct preference when trained on plus of 53%



217 (average result of 20 simulated bees) and just 52% for the reciprocal cross protocol. These simulations used  
218 the model parameters obtained from the bumblebee experiments: average flight speed whilst scanning of  
219 0.1m/s, and a distance from the stimuli of 20mm. However, when the experiment was reconfigured for the  
220 simulated bees to scan only the bottom half of the patterns, or just the lower left corner (as seen with the real  
221 bees (MaBouDi et al., 2021), the correct choice performance increased to  $\geq 96\%$  and  $\geq 98\%$  respectively (Figure  
222 2C). Increasing the scanning speed (*i.e.* increasing the separation between image patches) reduced the average  
223 performance to 70%. Similarly, simulated stationary bees only achieved 60% correct choices.

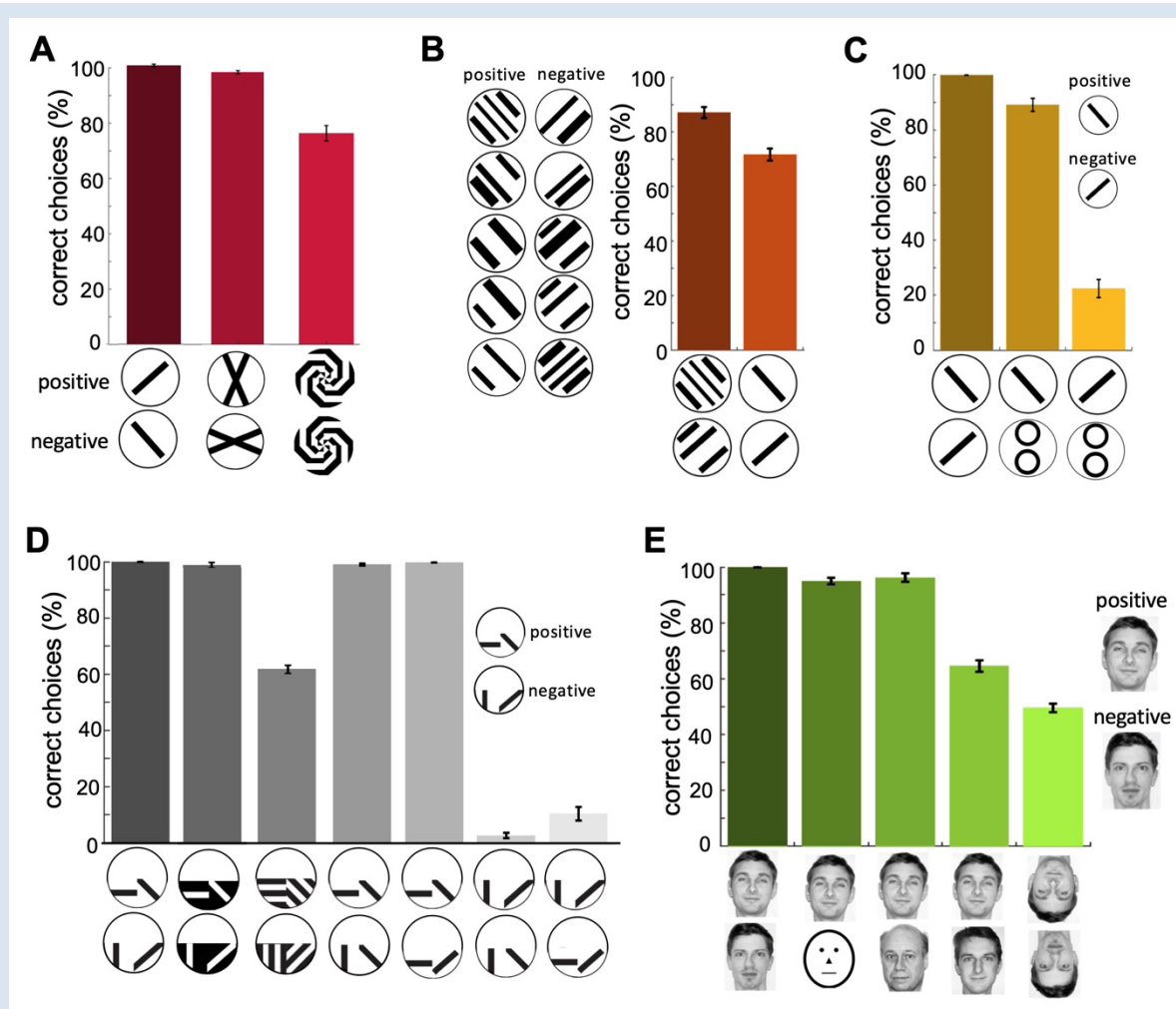
224

### 225 ***Neural network model of active vision bee behaviours in a variety of visual experiments***

226 In this study, we evaluated our model (utilising scans from the lower half only) by comparing it with other  
227 bees' experiments reported in the literature. It is important to note that bees may exhibit variations in  
228 scanning behaviour under different patterns and training conditions (see Discussion section). We found our  
229 simulated bees could discriminate angled bars (Hateren et al., 1990), a 22.5° angled cross from a 90° rotated  
230 version (Srinivasan, 1994) and spiral patterns (Zhang and Horridge, 1992) (Figure 3A). If trained on five  
231 perpendicular grating patterns, the simulated bees correctly identified the correct novel grating and a single  
232 bar pattern variant (Figure 3B). Figure 3C shows that not only could the proposed neural network learn to  
233 identify the correctly oriented bar pattern, but also identify the rewarding pattern from a novel one (two  
234 circles). More importantly the model showed a lower preference (22%) for the negatively trained pattern to  
235 that of the same novel pattern. This validates the implementation of the rejection behaviour in the model,  
236 demonstrating that the model simultaneously learns the rewarding and aversive stimuli. This was further  
237 explored by training the neural networks with patterns that contained two oriented bars in each lower  
238 quadrant of the patterns (Figure 3D) (Benard et al., 2006; Stach et al., 2004; Zhang and Horridge, 1992). The  
239 simulated bees again discriminated the training patterns without difficulty (over 99% accuracy). However, they  
240 performed worse on a simplified variant of the patterns with an average result of just 61%. When tested with  
241 the original positive pattern and novel patterns that contained just one correct orientation the bees had a high  
242 preference for the correct stimulus. Equally, the simulated bees showed a preference, if not as dominant, for  
243 a pattern that contained just one correct feature versus the trained negative pattern; showing that the model  
244 was able to extract more than a single feature during its scan of the pattern. To provide a substantially more  
245 complex pattern recognition task, we replicated the facial recognition experiment performed on honeybees  
246 (Dyer et al., 2005). We trained the neural network with images of two human faces (Figure 3E). Similar to the  
247 performance seen in honeybees, our simulated bees were able to identify the positive trained face from the  
248 negative one, two novel faces and a caricature of a face. Both the real bees and our simulated ones were  
249 unable to discriminate the faces when rotated through 180°. This result shows that complex visual features  
250 can be condensed through spatiotemporal encoding in the lobula neurons into specific and distinct neuronal



251 representations needed for learning in a miniature brain of the bee. This result is quite remarkable but shows  
 252 that complex visual features can be condensed through spatiotemporal encoding in the lobula neurons into  
 253 specific and distinct neuronal representations needed for learning in a miniature brain of the bee.  
 254



**Figure 3. Minimal neural network performance to published bee pattern experiments.** Twenty simulated bees, with random initial neuronal connectivity in mushroom bodies (see Methods), were trained to discriminate a positive target pattern from a negative distractor pattern (50 training exposures per pattern). The simulated bees' performances were examined via unrewarded tests, where synaptic weights were not updated (average of 20 simulated pattern pair tests per bee). Except for (A) all simulations were conducted at the default distance (2cm) and normal speed (0.1m/s) scanning the lower half of the pattern. **(A)** Mean percentage of correct choices ( $\pm$ SEM) in discriminating bars oriented at 90° to each other, 25.5° angled cross with a 45° rotated version of the same cross, and a pair of mirrored spiral patterns (MaBouDi et al., 2021b; Srinivasan, 1994). The simulated bees achieved greater than chance performances. **(B)** Performance of simulated bees trained with a generalisation protocol (Benard et al., 2006). Trained to 6 pairs of perpendicular oriented gratings (10 exposures per grating). Simulated bees then tested with a novel grating pair, and a single oriented bar pair. The simulated bees performed well in distinguishing between the novel pair of gratings; less well, but still significantly above chance, to the single bars. This indicates that the model can generalise the orientation of the training patterns to distinguish the novel patterns. **(C)** Mean performance ( $\pm$ SEM) of the simulated bees in discriminating the positive orientation from negative orientation. Additionally, the performance in recognising the positive

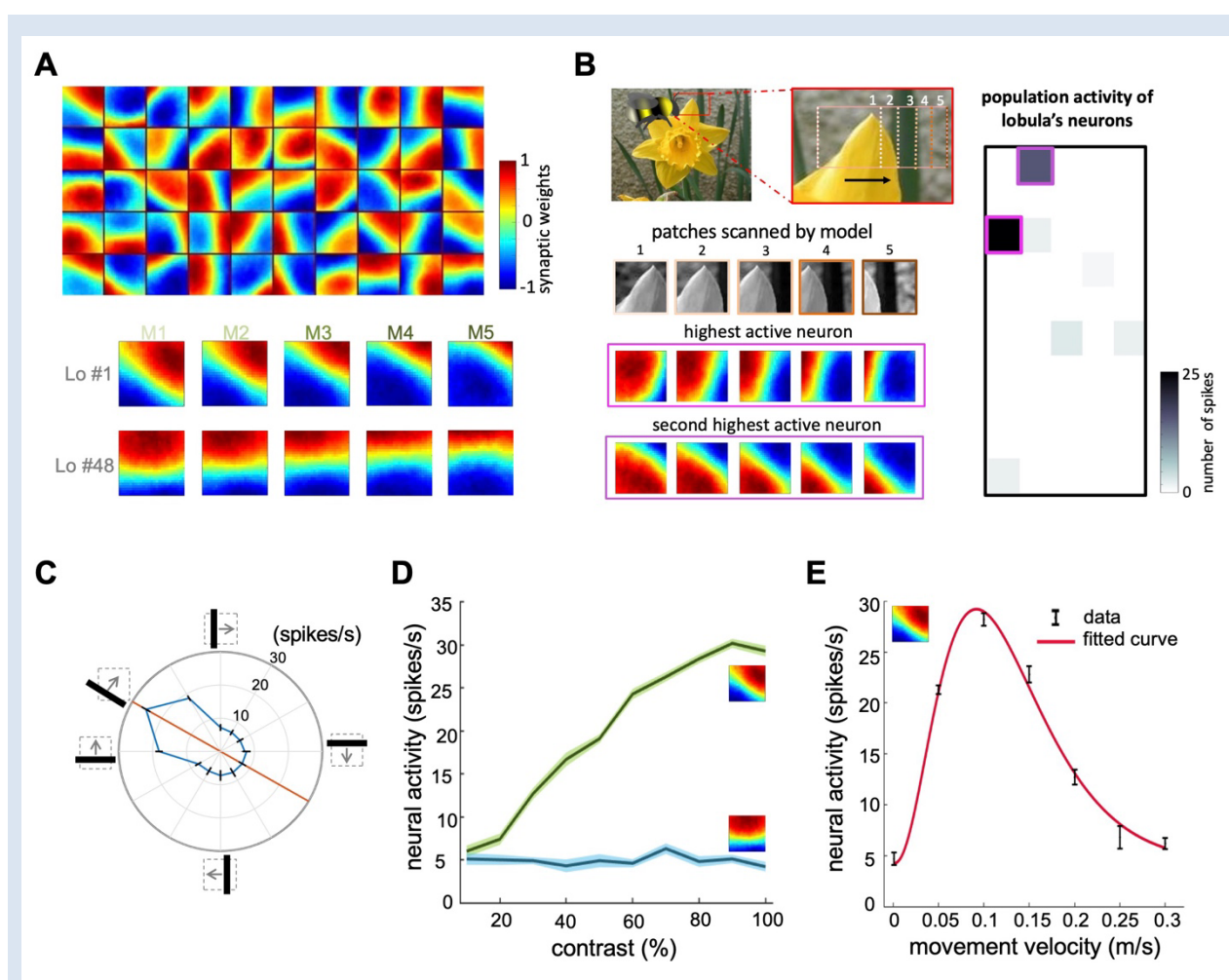
orientation from the novel pattern, and preference for the negative pattern from a novel pattern. Simulated bees learnt to prefer positive patterns, but also reject negative patterns, in this case preferring novel stimuli. **(D)** Performance of simulated bees trained to a horizontal and  $-45^\circ$  bar in the lower pattern half versus a vertical and  $+45^\circ$  bar (Stach et al., 2004). The simulated bees could easily discriminate between the trained bars, and a colour inverted version of the patterns. They performed less well when the bars were replaced with similarly oriented gratings, but still significantly above chance. When tested on the positive pattern vs a novel pattern with one correctly and one incorrectly oriented bar the simulated bees chose the positive patterns (fourth and fifth bars), whereas with the negative pattern versus this same novel pattern the simulated bees rejected the negative pattern in preference for the novel pattern with single positive oriented bar (two last bars). **(E)** The graph shows the mean percentage of correct choices for the 20 simulated bees during a facial recognition task (Dyer et al., 2005). Simulated bees were trained to the positive (rewarded) face image versus a negative (non-rewarded) distractor face. The model is able to recognise the target face from distractors after training, and also to recognise the positive face from novel faces even if the novel face is similar to the target face (fourth bar). However, it failed to discriminate between the positive and negative faces rotated by  $180^\circ$ .

255

### 256 ***Neural network response results during pattern learning***

257

258 The lobula neurons were additionally configured to laterally inhibit each other, allowing for non-associative  
259 learning (synaptic plasticity in the absence of reward) by employing synaptically local rules between lamina,  
260 medulla and lobula neurons (see Method section). The feed-forward weights from lamina to medulla neurons  
261 were updated with Oja's implementation of Hebb's rule (Oja, 1982) while symmetric inhibitory spike-timing-  
262 dependent plasticity (iSTDP) rule was implemented in synaptic connectivity of the lateral inhibitory lobula  
263 neurons (Vogels et al., 2011). These rules drive the network to efficiently represent the visual input with a  
264 limited number of lobula neuron activations (Figure 4B) (see Discussion). Figure 4A illustrates the obtained  
265 receptive fields of lobula neurons representing the spatiotemporal orientation-selectivity obtained after  
266 training with 100 flower and natural images (50,000 time-varying image patches). Each square represents one  
267 of the 50 lobula neurons, with the heat map representing the synaptic weights of the associated lamina  
268 neurons (connected via the medulla neurons). As representing spatiotemporal results in Figure 4A is inherently  
269 unintuitive, an example of two lobula neurons is shown below the matrix with the lamina synaptic weights for  
270 each of the five medulla neurons (see temporal dynamic of obtained RFs in Video 3 for all 50 lobula neurons,  
271 and Figure 4 - figure supplement 1 and Video 4 for all 100 lobula neurons). The receptive fields of a lobula  
272 neuron contain an elongated 'on' area (positive synaptic weights), next to an antagonistic "off" area (negative  
273 synaptic weights) that are mostly arranged in parallel in a specific orientation. The proportion of both 'on' and  
274 'off' areas continuously change from one time-delayed instance of medulla responses to the next. In the first  
275 example, the lobula cell with such a receptive field responds best to a  $135^\circ$  angled bar moving in the direction  
276 orthogonal to the orientation of the 'on' or 'off' areas and produces little or no responses to other orientations  
277 (Figure 4C). Our population of 50 lobula neurons show specificity to orientation and direction, similar to the  
278 neuronal responses observed in the brain of bees and other insects (James and Osorio, 1996; Paulk et al.,  
279 2008; Seelig and Jayaraman, 2013; Yang and Maddess, 1997).



**Figure 4. Neural responses of the simulated bee model to visual patterns. (A)** Top: each square in the matrix corresponds to a single time slice of the obtained spatiotemporal receptive field of a lobula neuron (5x10 lobula neurons) that emerged from non-associative learning in the visual lobes after exposing the model to images of flowers and nature scenes (see Video 3). Bottom: spatiotemporal receptive field of two example lobula neurons are visualised in the five-time delay slices of the matrices of synaptic connectivity between lamina and five medulla neurons (See Figure 1D). The lobula neuron integrates signals from these medulla neurons at each of five time periods as the simulated bees scan a pattern (time goes from left to right). Blue and red cells show inhibitory and excitatory synaptic connectivity, respectively. The first example lobula neuron (#1) encodes the 150° angled bar moving from lower left to the upper right of the visual field. The second example lobula neuron (#48) encodes the movement of the horizontal bar moving up in the visual field. **(B)** An example of an image sequence projected to the simulated bee' eye with lateral movement from left to right. Below shows the five images patched sampled by the simulated bee. The right side presents the firing rate of all lobula neurons responding to the image sequence. The spatiotemporal receptive field of two highest active neurons to the image sequence are highlighted in purple. **(C)** The polar plot shows the average orientation selectivity of one example lobula neuron (#1) to differently angled bars moving across the visual field in a direction orthogonal to their axis (average of 50 simulations). This neuron is most sensitive to movement when the bar orientation is at 150°. **(D)** The spiking response of the lobula neuron to the preferred orientation raised as the contrast was increased, whereas the response of the lobula neuron to a non-preferred orientation is maintained irrespective of contrast. **(E)** The average velocity-sensitivity curve ( $\pm$ SEM) of the orientation-sensitive lobula neuron (#1) is obtained from the responses of the lobula neuron to optimal

(angle of maximum sensitivity) moving stimuli presented to the model at different velocities. The red line shows the Gamma function fitted to the data.

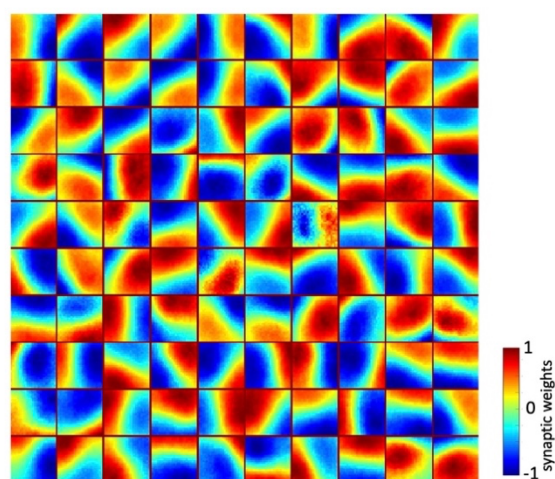
281

**Video 3: Temporal dynamic of receptive field of 50 lobula neurons obtained from non-associative learning procedure and active scanning.**

282

283 For clarity, Figure 4B shows how a sequence of image patches simulating a horizontal scan is processed, and  
284 results in a limited number of lobula neuron responses. This means that the activity of lobula neurons is  
285 decorrelated and relatively sparse as a result of non-associative learning mechanisms in the visual lobe (See  
286 Discussion). Interestingly, the receptive field of two first active lobula neurons captured the visual structure of  
287 the flower's petal that was scanned by the simulated bee (*i.e.* one is matched with left 45 angled edge of the  
288 flower's petal and the other is almost matched with the right angled edge of the flower's petal). This shows  
289 that our model can extract the different visual features of the input with a minimum number of filters (lobula  
290 neurons). To further demonstrate the characteristics of the 150° sensitive lobula neurons (as one example),  
291 the spiking activity of the neuron is calculated in the response to a set of oriented bars as they were moved in  
292 the direction orthogonal to their orientation. As expected, the neuron is maximally sensitive to a moving 150°  
293 bar (26 spikes/sec), it still responds to a horizontal bar and a moving 120° bar (18Hz) but has limited responses  
294 (above base firing rate) to the other stimuli (Figure 4C). Consistent with neural observations (Yang and  
295 Maddess, 1997), the firing rate response of our lobula neurons, at their 'preferred' orientations, rises as the  
296 contrast is increased. Conversely, their responses to the non-preferred orientations are constant irrespective  
297 of contrast (Figure 4D). Interestingly, these lobula neurons are also velocity-sensitive (Figure 4E). Thus, each  
298 lobula neuron optimally responds to a specific orientation and velocity. This demonstrates that our model  
299 captures many quantitative features of lobula neurons (Paulk et al., 2008; Yang and Maddess, 1997).

300



**Figure 4 - figure supplement 1:** The spatiotemporal receptive field of lobula neurons emerged from the non-associative learning if the number of lobula neurons is set to 100 (see Video 4).

301

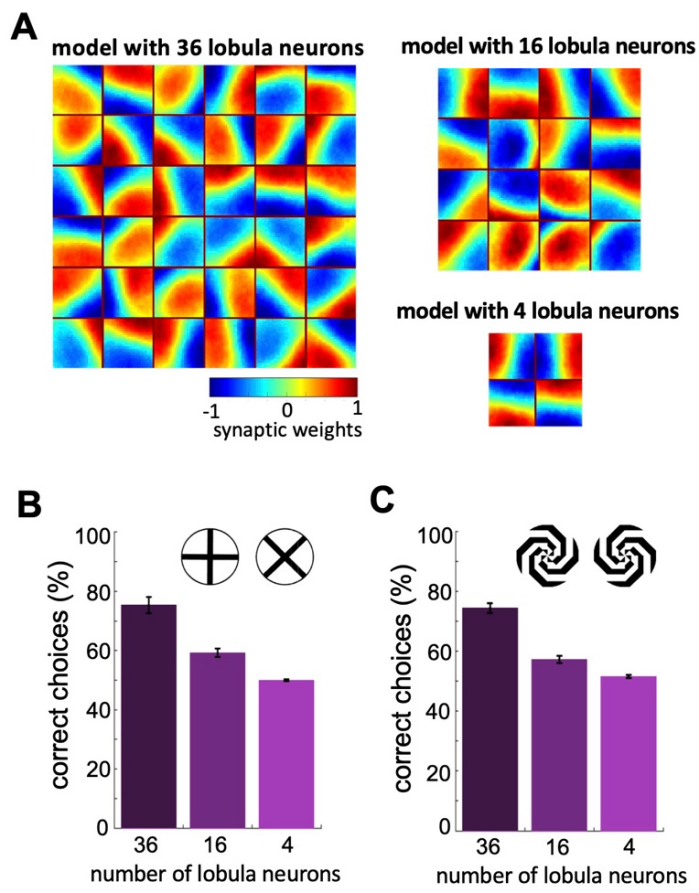
**Video 4: Temporal dynamic of receptive field of 100 lobula neurons obtained from non-associative learning procedure and active scanning.**

302

303 **What is the minimally sufficient circuitry required for active vision in bees?**

304 As reported above, the model is able to accomplish a variety of pattern recognition tasks (Figures 2, 3). We  
305 asked whether our neural networks could perform with a very limited number of lobula neurons that transfer  
306 visual information to the mushroom body. Hence, we conducted the non-associative learning process with  
307 varying number of lobula neurons, setting them to either 4, 16, or 36 (the original model had 50 lobula  
308 neurons). Then the visual network was trained using the same set of natural images and protocol defined for  
309 the original model. Interestingly, the non-associative learning process led to the emergence of distinct spatio-  
310 temporal structures in the lobula neurons. We found that the variability between the spatiotemporal receptive  
311 field of lobula neurons is reduced by decreasing the number of lobula neurons (Figure 5A and Videos 5, 6 ,7).  
312 The model cannot encode the spatio-temporal structure inherent in the training patterns if the model is  
313 limited to 4 neurons, only vertical and horizontal receptive fields were created (Video 5). As expected, the  
314 performance of the model decreased by reducing the number of lobula neurons. However, the model with 16  
315 lobula neurons can still solve the discrimination between plus and multiplication signs, and solve a difficult  
316 visual task above chance (Figure 5B, C). This finding highlights how the spatio-temporal encoding in the visual  
317 lobe enhances the model's capacity to represent the visual environment using fewer neurons, compared to  
318 what is typically necessary in a minimal model.





**Figure 5. Minimum number of lobula neurons that are necessary for pattern recognition.**

**(A)** Obtained spatiotemporal receptive field of lobula neurons when the number of lobula neurons were set at 36, 16 or 4 during the non-associative learning in the visual lobe (See Figure 5A). This shows the models with lower number of lobula neurons encode less variability of orientations and temporal coding of the visual inputs (see Videos 5,6) **(B & C)** The average correct choices of the three models with 36, 16 or 4 lobula neurons after training to a pair of plus and multiplication patterns (B) and mirrored spiral patterns (C). The model with 16 lobula neurons still can solve pattern recognition tasks at a level above chance. It indicates that only 16 lobula neurons that provide all inputs to mushroom bodies are sufficient for the simulated bees to be able to discriminate between patterns.

319

320

**Video 5: Temporal dynamic of receptive field of 4 lobula neurons obtained from non-associate learning procedure and active scanning (compare it to Video 3).**

321

**Video 6: Temporal dynamic of receptive field of 16 lobula neurons obtained from non-associate learning procedure and active scanning (compare it to Video 3).**

322

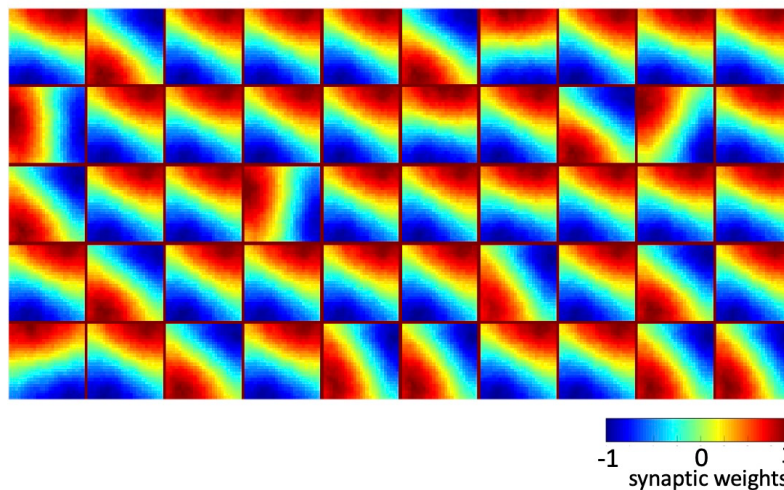
**Video 7: Temporal dynamic of receptive field of 36 lobula neurons obtained from non-associate learning procedure and active scanning (compare it to Video 3).**

323

324 Moreover, to explore the effect of inhibitory neurons within the visual lobe on the output of lobula neurons,  
325 the model was trained using the same protocol, but the synaptic weights of the inhibitory connections to the  
326 lobula neurons were not updated during exposure to the training images. These fixed inhibitory connections



327 caused a limitation in shaping the population of lobula neurons to encode moving orientations (Figure 6 and  
328 Video 8). It indicates that the presence of inhibitory interneurons in the visual lobe plays a crucial role in  
329 facilitating an efficient representation of the visual environment.  
330



**Figure 6. The role of lateral inhibitory connections between lobula neurons.** Obtained spatiotemporal receptive field of lobula neurons when the lateral inhibitory connectivity between lobula neurons is fixed during the non-associative learning (see Video 8 and Figure 4A & Video 3).

331

**Video 8: Temporal dynamic of receptive field of 50 lobula neurons when the lateral inhibitory connectivity between lobula neurons is fixed during the non-associative learning (compare it with Video 3)**

332

333 Taken together, these findings demonstrate that our assumption regarding non-associative plasticity in the  
334 visual lobe can successfully replicate the neural responses of lobula neurons across various patterns and  
335 conditions. This results in a sparse and uncorrelated representation of the visual input, which is advantageous  
336 for subsequent learning processes in the mushroom body. Importantly, these results align closely with  
337 theoretical studies (see Discussion section), further supporting the effectiveness of the active vision in  
338 capturing the underlying principles of information encoding in the insect visual system.

339

## 340 Discussion

341 In this study, we aimed to gain insights into the computational requirements for visual pattern recognition by  
342 investigating a minimal neural network. To achieve this, we leveraged the flight behaviours of bees during  
343 their active scanning of visual patterns and developed a novel model inspired by the insect's visual system.  
344 Through simulations, we examined how the visual environment is represented through the spatio-temporal  
345 responses of a small population of neurons in the lobula region in the visual lobe. By incorporating non-

346 associative learning into our model, we discovered that it can effectively shape the connectivity within the  
347 visual lobe and generate an efficient representation of the input. This self-organization process led to the  
348 emergence of orientation-selective cells in the lobula, which played a vital role in encoding the complex visual  
349 environment. Our model of the bees' visual system, and subsequent simulations, demonstrate that the  
350 complex visual environment can be condensed into spatiotemporal representations, expressed through the  
351 firing rate responses of a small population of lobula neurons sensitive to specific orientations and velocities.  
352 Not only are these limited representations capable of discriminating the plus and multiplication patterns used  
353 in our behavioural experiments (MaBouDi et al., 2021b), but can also generalise from trained patterns to novel  
354 stimuli, and even provide accurate results in a task of human face recognition, indicating its potential for  
355 broader applicability. Our findings also suggest that the movement of bees, or their active vision, may play a  
356 crucial role in their ability to efficiently analyse and encode their environment. The spatio-temporal encoding  
357 within the visual lobe appears to be a key mechanism employed by bees to achieve this efficient information  
358 representation. Overall, our study sheds light on the necessary computational requirements for visual pattern  
359 recognition, highlighting the significance of active vision and spatio-temporal encoding within the insect's  
360 visual system. These insights have implications not only for understanding the information processing  
361 capabilities of bees but also for inspiring the development of novel computational models for visual  
362 recognition tasks.

363 The question of how animals deal with noisy and complex natural world has a long history (Barlow, 1961;  
364 Gibson, 1979; Menzel and Giurfa, 2006; Srinivasan, 2010). The efficient coding hypothesis is one of the  
365 important theoretical hypotheses in this field that refers to the phenomenon that the early visual system  
366 compresses inputs into a more efficient form to transmit relevant visual information with maximum  
367 information to higher order brain regions (Barlow, 1961). According to this hypothesis, the responses of a  
368 single cell in the visual system to the natural environment should entirely utilise its output capacity (e.g.,  
369 maximum firing rate) and the population responses of different neurons to the natural signals should be  
370 statistically independent (Simoncelli and Olshausen, 2001). Bees present a wide range of complex behaviours  
371 over small scales of inspections and large scales of navigation (Menzel, 2012; Srinivasan, 2010) despite their  
372 limited computational resources. Thus, bees are an appropriate model to explore the effect of ecological  
373 constraints in the neural computation underlying cognition. The non-associative model presented in this study  
374 supports the efficient coding hypothesis (Figures 4 & 5). The suggested model works as a linear generative  
375 model that successfully replicates the receptive fields of cells in the lobula (Barlow, 1961; Olshausen, 2003).  
376 After training, the correlation among activity of lobula neurons to spatiotemporal naturalistic signals is highly  
377 reduced. Further, only a limited number of lobula neurons respond to specific visual stimuli (Figure 4). This  
378 indicates that the model removes redundancy in the natural scenes such that the receptive fields of lobula

379 cells often encode perceptually salient features in the natural scene and transfer the incoming signals into a  
380 more efficient form.

381

382 In this study, we use the type of natural scenes with a statistical structure similar to those that visual systems  
383 have adapted to over evolutionary time periods (Geisler, 2008; Hyvärinen et al., 2009; Simoncelli and  
384 Olshausen, 2001). Also, since the main goal of bee navigation and foraging is finding food from a large variety  
385 of potential flower resources, our non-associative network was trained with a set of different flower images.  
386 As with all theoretical models this is a simplification, as real bees traverse a 3D environment viewed through  
387 a 270° field of view. Here, we assume that formation of receptive fields would be equivalent to that of our 2D  
388 simulations. However, further studies are necessary to refine and expand our model based on a more  
389 comprehensive understanding of the function and structure of the bee eye components (Juusola et al., 2017;  
390 Taylor et al., 2019). Moreover, investigating the neural mechanisms underlying visual learning in the bee brain  
391 will allow us to fine-tune our model's architecture and parameters, leading to a more faithful representation  
392 of the bee visual system. Through continued research and collaboration, we plan to refine our model and gain  
393 deeper insights into the remarkable capabilities of the bee's visual perception.

394

395 Our findings align with previous studies on bumblebees' discrimination of plus and multiple sign patterns  
396 (MaBouDi et al., 2021b), indicating improved performance of our model when focusing on scanning the lower  
397 half of patterns at specific velocities. However, bees exhibit variations in scanning behaviour under different  
398 patterns and training conditions (Giurfa et al., 1999; Guiraud et al., 2018). It has been shown that honeybees  
399 and bumblebees can solve visual tasks by extracting localized or elemental features within patterns, adapting  
400 their discrimination strategies accordingly (Giurfa et al., 1999; MaBouDi et al., 2021b; Stach et al., 2004; Stach  
401 and Giurfa, 2005). Recent analysis of bee flight paths supports these findings, revealing how honeybees can  
402 successfully solve tasks by selectively scanning specific elements in stimuli (Guiraud et al., 2018). This suggests  
403 that bees may develop tailored flight manoeuvres during training, optimizing their scanning behaviour to  
404 extract maximum visual information based on patterns and protocols. Although our model highlights the  
405 importance of studying active vision, it is essential to recognize that further investigations are needed to  
406 explore the optimal flight scanning behaviour in bees. Additionally, incorporating an adaptive vision-motor  
407 loop into our models will enable a more comprehensive understanding of active vision in insects. These  
408 advancements will provide valuable insights into how insects perceive and interact with their visual  
409 environment, ultimately enhancing our understanding of the mechanisms underlying active vision.

410

411 The results of our model suggest that passive visual exposure to natural images modifies the connectivity in  
412 the visual lobes and leads to better ability in pattern recognition (Figures 2, 3). These developed synaptic

413 connections emerged independent of the initial connectivity that each simulated bee had upon creation. We  
414 assume that the specific visual experiences that real bees are exposed to in early life define the individual  
415 capabilities of each bee's visual representation, which may influence their later performance in behavioural  
416 tasks (Hertel, 1983, 1982; MaBouDi et al., 2017; Vetter and Visscher, n.d.). There is direct empirical evidence  
417 for such neural developmental processes in olfactory systems of bees, where early passive exposure improved  
418 the bees' later ability in odour discrimination (Arenas and Farina, 2008; Locatelli et al., 2013). Our previous  
419 research of olfactory coding demonstrated that the iSTDP learning rule can create specific connectivity in the  
420 sensory system and increase the separability of odour representations in the antennal lobe outputs (MaBouDi  
421 et al., 2017). A similar mechanism was developed here between lobula neurons, such that only a limited  
422 population of lobula neurons are excited for particular visual inputs, providing sparse and distinct outputs to  
423 the mushroom body learning centres (Figure 3B). The obtained receptive fields of lobula cells with the fixed  
424 lateral connectivity shows the inhibition is required for orientation selectivity and temporal coding in the visual  
425 lobe (Fisher et al., 2015). Our results highlight the important functions of inhibitory connections within the  
426 visual lobes. Accordingly, our model predicts that bees with less experiences of visual processing at the early  
427 stage of life are worse in the learning and memory of visual tasks compared with bees with rich visual  
428 experiences. However, further behavioural and neurobiological studies are required to assess this prediction.

429

430 Mushroom bodies are critical centres for associative learning and memory in insects (Heisenberg, 2003;  
431 Menzel, 2012). Synapses between Kenyon Cells and extrinsic mushroom body neurons obey a Hebbian STDP  
432 rule (Cassenaer and Laurent, 2007; Markram et al., 1997). However the STDP rule alone cannot maintain  
433 associative learning (Abbott and Nelson, 2000; Meeks and Holy, 2008). Associative learning in insects appears  
434 to rely on the neurotransmitters octopamine and dopamine, to reflect unconditioned signals for appetitive  
435 and aversive valances (Cognigni et al., 2018; Hammer, 1993; Hammer and Menzel, 1995; Perry and Barron,  
436 2013; Schwaerzel et al., 2003). These are released into the mushroom body lobes where Kenyon cells connect  
437 to MB output neurons (MBON) (Burke et al., 2012; Okada et al., 2007; Strube-Bloss et al., 2011). Cassenaer  
438 and Laurent, using in-vivo electrophysiology in locusts, reported the depressive action of octopamine on  
439 synapses underlying STDP rule that leads to a lower response for MBONs in the presence of octopamine  
440 (Cassenaer and Laurent, 2012). Hence, following this observation, we model associative learning of pairing the  
441 positive pattern with the reward by the octopamine modulation of STDP (Equation 4; Figure 7). Here, both  
442 temporal ordering of pre- or post- synaptic spikes depress the synaptic connection between Kenyon cells and  
443 the MBON.s Also, the synapses are updated when the negative patterns are paired with the punishment  
444 following the classical STDP (Equation 3; Figure 7). This combination results in a complex interplay between  
445 synaptic changes and reinforcer signals and enriches the model to not only learn to correctly choose the  
446 positive patterns, but also learn to reject incorrect patterns (Figure 2B, 3C). The changes to the synaptic weight

447 of connections results in a decreasing response of MBON to the positive patterns during the associative  
448 learning which is consistent with the PE1 extrinsic neuron in the honeybee brain that exhibits a lower response  
449 to the positive patterns (Okada et al., 2007) (Figure 2B). However, further studies are required to investigate  
450 the novel combination of octopamine and dopamine modulation of STDP that is introduced in this study.  
451 Together the non-associative learning in the optic lobes and supervised learning in the mushroom bodies  
452 produced a model capable of not only discriminating simple patterns but also generalisation (Figure 3B), and  
453 correct judgments in conflicting stimulus experiments (Figure 3D). However, the real power of this approach  
454 is exemplified in the facial recognition task (Figure 3E). Here, the complexity of the human face is reduced to  
455 a number of sparse lobula neuron activations that can be learnt by the mushroom bodies. But more interesting  
456 is that the spatiotemporal receptive fields formed during non-associative learning respond differently for  
457 different faces, allowing fine differences to still be sufficiently encoded. Real bees rarely have to discriminate  
458 between human faces, but these same processes undoubtedly aid bees in selecting rewarding flowers without  
459 requiring a complex visual memory within their miniature brains.

460

461 Recent studies showed that bees can sometimes use a more efficient, less-cognitively demanding strategy to  
462 solve a cognitive task (Cope et al., 2018; Guiraud et al., 2018; Langridge et al., 2021; MaBouDi et al., 2023,  
463 2021a, 2020b; Roper et al., 2017; Vasas et al., 2019). Most of bees' responses to different cognitive tasks can  
464 be described by a simple neural network. For instance, Roper et al. (2017) suggest that reliable generalisation  
465 of visual information can be achieved through simple neuronal circuitry that is biologically plausible and can  
466 be accommodated in a small bee brain (Roper et al., 2017). Also, a mathematical model of colour processing  
467 in the bee brain propose that the diversity of colour-sensitive responses can be explained using a simple model  
468 by the assumption that these neurons receive randomly weighted inputs from all receptor types, and that this  
469 type of neural organisation is likely implemented during neural development and experience-dependent  
470 manner (MaBouDi et al., 2020b; Vasas et al., 2019). Together, the behavioural and computational experiments  
471 developed during our research emphasise the fundamental roles of exploring the mechanisms of cognitive  
472 abilities in animals with miniature brains and designing minimum neural networks to understand the  
473 requirements of certain cognitive tasks. A mechanistic investigation of how bees parse natural environments  
474 provides basic principles for current challenging problems in designing autonomous robots. Indeed, these  
475 computational shortcuts that have evolved for billions of years will enable us to develop more efficient  
476 artificial intelligence, capable of solving specific problems much more effectively than humans and current  
477 artificial intelligence (de Croon et al., 2022; Webb, 2020).

478

## 479 **Materials and Methods**

### 480 ***Network topology of active vision model***

481 The model architecture of the bee visual pathway is shown in Figure 1. A bumblebee has a pair of compound  
482 eyes that are composed of ~5,500 ommatidia (Spaethe and Chittka, 2003; Streinzer et al., 2013). Each eye  
483 contains three different types of photoreceptors, short, medium and long wavelength sensitive peaking in the  
484 UV, blue and the green respectively (Menzel and Blakers, 1976; Skorupski et al., 2007). Since the green  
485 photoreceptors are those that predominantly mediate visual pattern recognition (Giger and Srinivasan, 1996;  
486 Spaethe et al., 2001), we modelled that 75x75 green photoreceptors in one eye component are activated by  
487 the pixel values of the input pattern. Photoreceptors then project to 625 (25x25) neurons in the lamina, which  
488 is the first centre of visual processing. In this model, one lamina neuron,  $r_l^{La}$ , receives and sums inputs from  
489 nine neighbouring photoreceptors placed in neighbouring ommatidia, as  $r_l^{La} = f(\sum_{p=1}^P r_p; a, b)$ . Here,  
490  $f(x; a, b) = A_0 / (1 + \exp(mx + b))$  is the activation function where  $A_0 = 1$  is the maximum activity of the  
491 lamina neurons and parameters  $m = 1$  and  $b = 0$  control the shape of the activation function  $f$ .

492 Each medulla neuron is activated by the summation of activity of the lamina neuron via the synaptic  
493 connectivity  $W$ . Each spiking neuron in this study follows the integrate-and-fire model. The dynamics of the  
494 subthreshold membrane potential of a neuron,  $u(t)$  is described by the following standard conductance-  
495 based leaky integrate-and-fire model:  $\tau \frac{du(t)}{dt} = -u(t) + R \cdot I(t)$ , where  $R = 10$  and  $\tau = 10ms$  are the  
496 resistance and membrane time constant of the neuron respectively. Here, the input  $I(t)$  exhibit the total  
497 synaptic input to the cell from presynaptic neurons.

498 The membrane potential is reset to the base activity,  $v_0 = -80 mV$ , if it exceeds the threshold,  $V_T = 0 mV$ .  
499 However, the input of the  $m$  –the medulla neuron is calculated  $I_m^{Me} = \sum_{l=1}^L W_{m,l} r_l^{La}$ . The value  $W_{l,m}$  specifies  
500 the strength of a synaptic input from the  $l$  –th lamina neuron to the  $m$  –the medulla neuron. To model the  
501 variability of neural responses, a signal noise generated by Poisson distribution was added to the output of  
502 the neuron.

503

504 We propose a temporal coding model between the medulla and lobula neurons. Each wide field lobula neuron  
505 receives synaptic input from  $M$  small field medulla neurons with delay  $T$  (Figure 1D) ( $M$  corresponds to the  
506 number of frames in the input of the model). These small field medulla neurons are activated from  $M$  different  
507 regions of the image patch via the pathway passing through the photoreceptors and lamina neurons (Figure  
508 3B). While each medulla neuron is affected by  $1/M$  of the pattern, a lobula neuron is activated by the whole  
509 pattern.  $M$  medulla neurons that are sampled from a selected region with delay  $T_0$  show the neural  
510 representation of the bees' scanning behaviour in front of the pattern. The medulla neurons send their spiking  
511 responses to a wide-field lobula neuron with a synaptic delay such that all signals activate the lobula neuron  
512 in the same instance. The parameters of the model's scanning behaviour (viewing distance: 2cm, flight speed  
513 0.1m/s) are obtained from bees whilst inspecting stimuli (MaBouDi et al., 2021b).



514 The lobula neurons are laterally interconnected by the inhibitory neurons via the synaptic connectivity  $Q =$   
515  $[q_{i,j}]$ , where  $q_{i,j}$  represents the lateral connectivity between  $i$  –th and  $j$  –th lobula neurons. This connectivity  
516 along with  $W_{m,l}$  are updated during a non-associative learning process (see next subsection). The neurons in  
517 the lobula region send excitatory signals to randomly selected Kenyon cells (KCs) in the mushroom bodies  
518 through the synaptic connectivity matrix (Caron et al., 2013; Szyszka et al., 2005)  $S = [s_{o,k}]$ . The positive  
519 values  $s_{o,k}$  exhibits the strength synaptic weight from  $o$ -th lobula neuron to  $k$ -th KC. KCs in the network have  
520 sparse activity, meaning they are selective to particular image features (i.e. each pattern activates less than  
521 5% of Kenyon cells(Honegger et al., 2011)). All Kenyon cells project to a single mushroom body Output Neuron  
522 (MBON), which is the final output of the model. The input of the of the MBON,  $I_{MBON}$ , is computed by the KC-  
523 MBON connections  $D$  such that  $I_{MBON} = \sum_{k=1}^K D_k r_k^{KC}$ , where  $r_k^{KC}$  is the spiking activity of the  $k$ -th KCs. Finally,  
524 a reinforcement neuron makes reinforcement-modulated connections with the KCs and MBON in the presence  
525 of the positive and negative patterns (see the next section).

526

### 527 ***Training the network via a non-associative learning***

528 We trained the model on 50,000 time-varying patches randomly selected from 100 flowers and natural scenes.  
529 At each step of training, a set of five patches with size 75x75 pixels, selected by shifting 15 pixels over the  
530 image from the left or right or the reverse orientation (Figs 1B, 2A), was considered as the input of the model;  
531 this would correspond to the process of bees' scanning part of the image. Using the network described above,  
532 the number of spikes from each lobula neuron are counted separately for each set of patches scanned through  
533 each movement. We start the training with all inhibitory connection strengths  $Q$ , where its elements are  
534 randomly generated from a uniform distribution between 0 and 1. The feed-forward synaptic weights are  
535 initialized with Gaussian white noise  $\mathcal{N}(0, 1)$ . The neural responses random time-varying patches evoked by  
536 the images were used to update the connections strengths  $Q$  and  $W$ , simultaneously (see Discussion section).

537 After the image presentation, the feed-forward weight  $W$  is updated according to Oja's implementation of the  
538 Hebbian learning rule (MaBouDi et al., 2017; Oja, 1982) via

$$539 \quad \Delta W_{ij} = \gamma r_j^{Me} (r_i^{La} - r_j^{Me} W_{ij}) \quad (\text{Equation 1})$$

540 Here, the  $r_j^{Me}$  and  $r_i^{La}$  represent the activities of the  $j$ -th medulla and  $i$ -th lamina neurons, respectively. The  
541 positive constant  $\gamma$  defines the learning rate.

542 At the same time of processing, the lateral inhibitory connectivity in the lobula is modified by inhibitory spike-  
543 time-dependent plasticity (iSTDP) (Vogels et al., 2011). Here, we model *non-associative learning* in lobula by a

544 symmetric iSTDP between presynaptic of the inhibitory neurons and postsynaptic lobula neurons. In this  
545 learning rule, both temporal ordering of pre- or post- synaptic spikes potentiate the connectivity and the  
546 synaptic strength of j-th inhibitory neuron onto i-th lobula neuron ( $Q_{i,j}$ ) is updated as follows:

$$547 \quad \Delta Q_{i,j} = \eta(r_i^{Lo} * r_j^{In} - \alpha) \quad (\text{Equation 2})$$

548 where  $r_i^{Lo}$  and  $r_j^{In}$  exhibit the mean firing rate of the lobula and inhibitory neurons, respectively. The  
549 depression factor  $\alpha$  controls the target activity rate of the lobula neurons. Here,  $\eta$  is the learning rate. To  
550 simplify, a one-to-one connection between the inhibitory and lobula neuron is assumed in the model such that  
551 the activity of the j-th inhibitory neuron is equal to the activity of the j-th lobula neuron. The training is  
552 terminated when the synaptic weights over time are changed less than a very small threshold (0.001).

553

### 554 **Associative learning in Mushroom Bodies**

555 To verify if the lobula neurons can reproduce empirical behavioural results in different visual tasks, the model  
556 is enriched with a supervised learning process in the mushroom bodies. When the training process of the non-  
557 associative learning is terminated, we use a reward-based synaptic weight modification rule in KCs-MBON  
558 connection (D), such that, if a stimulus is rewarding (*i.e.* positive), the corresponding synapses between  
559 activated neurons will be weakened while for a stimulus paired with punishment (*i.e.* negative), activated  
560 synapses are strengthened (Cassenaer and Laurent, 2012) (see Discussion section). The model behaves as the  
561 activity of mushroom body neurons in decreasing their firing rate in responding to the positive stimuli during  
562 training (Okada et al., 2007). In this model, a single reinforcement neuron modulated strengths of synaptic  
563 connectivity at the output of the KCs in response to both reward and punishment. In the presence of the  
564 negative patterns, the synaptic strengths from the KCs to the MBON are modified, and modulated by  
565 dopamine, based on the classical STDP (Song et al., 2000; Zhang et al., 1998) (Figure S3A):

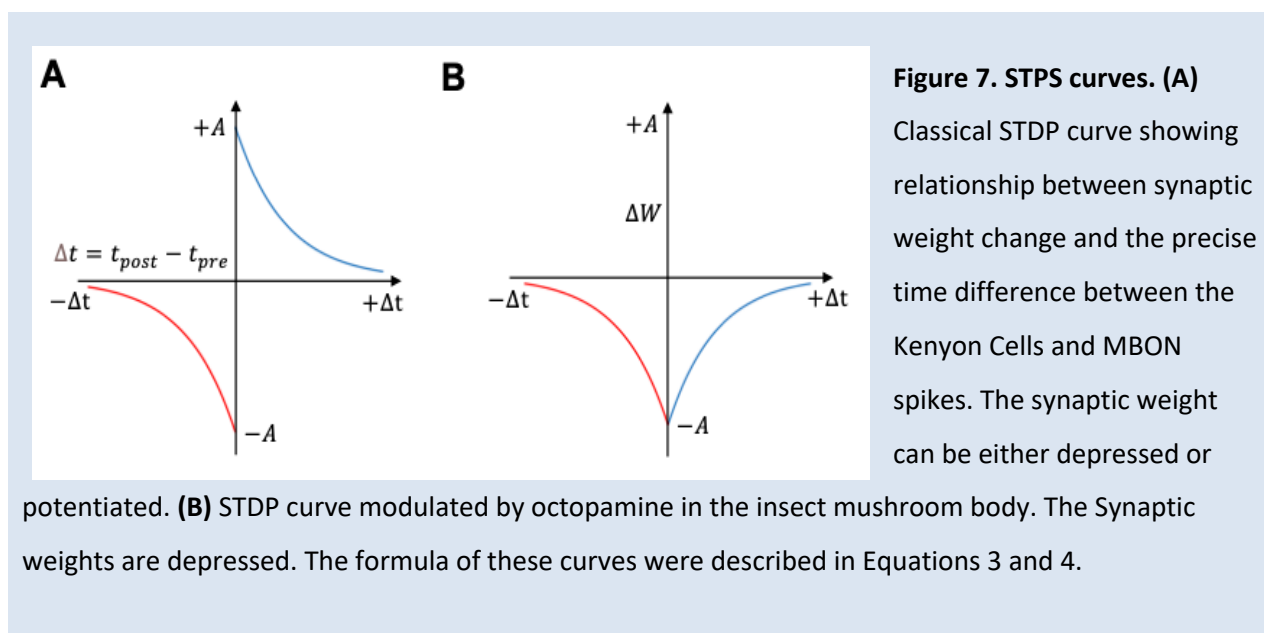
$$566 \quad STDP_{Dop}(\Delta t) = \begin{cases} A e^{-\Delta t/\tau}, & \Delta t > 0 \\ -A e^{\Delta t/\tau}, & \Delta t < 0 \end{cases} \quad (\text{Equation 3})$$

567 where  $\Delta t = t_{post} - t_{pre}$  implies the difference between the spike time of pre- and post- synaptic neurons.  
568 Further, applying the synaptic plasticity rule modulated by octopamine (octopamine modulated STDP)  
569 observed in the presence of rewarding stimuli to the synapses between KCs and MBON (Cassenaer and  
570 Laurent, 2012), the change in synaptic weight can be summarized as (Figure 7):

$$571 \quad STDP_{OCT}(\Delta t) = \begin{cases} -A e^{-\Delta t/\tau}, & \Delta t > 0 \\ -A e^{\Delta t/\tau}, & \Delta t < 0 \end{cases} \quad (\text{Equation 4}).$$

572 Here,  $A = 0.01$  and  $\tau = 20$  ms exhibit the maximum magnitude and time constant of the STDP function for  
573 the synaptic potentiation or depression.

574



575

576 To train the model in different conditions of scanning, the flight-scan forms of the positive and negative  
577 patterns were presented to the model. Each set of flight-scan input contained a set of five patches with size  
578 75x75 pixels were selected from the test patterns by shifting 15 pixels over each pattern from the left to right  
579 (Figure 2A). The numbers of shifted pixels control the speed of scanning. The activity of the MBON was used  
580 to assess the performance of the model. Following the training, the performance of the model was calculated  
581 from a decrease in firing rate of the MBON to a pattern that had been rewarding and/or an increase in firing  
582 rate of MBON to a pattern that had been punishing in training. The bee's final behavioural decision is proposed  
583 to come from a simple integration of these different valence-encoding neurons.

584

### 585 Acknowledgements

586 We thank Paul Graham and Andrew Barron for valuable comments on manuscript and Alice Bridges for  
587 drawing the front view of the bumblebee presented in Figure 1A. This research was financed by HFSP  
588 programme grant RGP0022/2014 and by EPSRC program grant Brains-on-Board EP/Poo6094/1.

589

### 590 Conflict of interest statement

591 All authors declare no conflict of interest.

592

## 593 **Data and code accessibility**

594 The code developed for this research project has been made openly accessible on GitHub:  
595 [https://github.com/hadiimaboudi/neuromorphic\\_model\\_active-vision-](https://github.com/hadiimaboudi/neuromorphic_model_active-vision-)

596

## 597 **References**

- 598 Abbott LF, Nelson SB. 2000. Synaptic plasticity: taming the beast. *Nat Neurosci* **3**:1178–1183.
- 599 Anderson AG, Ratnam K, Roorda A, Olshausen BA. 2020. High-acuity vision from retinal image motion. *J Vis*  
600 **20**:34–34.
- 601 Arenas A, Farina WM. 2008. Age and rearing environment interact in the retention of early olfactory  
602 memories in honeybees. *J Comp Physiol A* **194**:629–640. doi:10.1007/s00359-008-0337-z
- 603 Arenz A, Drews MS, Richter FG, Ammer G, Borst A. 2017. The temporal tuning of the Drosophila motion  
604 detectors is determined by the dynamics of their input elements. *Curr Biol* **27**:929–944.
- 605 Avarguès-Weber A, Dyer AG, Giurfa M. 2011. Conceptualization of above and below relationships by an  
606 insect. *Proc R Soc Lond B Biol Sci* **278**:898–905. doi:10.1098/rspb.2010.1891
- 607 Barlow HB. 1961. Possible principles underlying the transformation of sensory messages. *Sens Commun* **1**.
- 608 Benard J, Stach S, Giurfa M. 2006. Categorization of visual stimuli in the honeybee *Apis mellifera*. *Anim Cogn*  
609 **9**:257–270. doi:10.1007/s10071-006-0032-9
- 610 Boeddeker N, Mertes M, Dittmar L, Egelhaaf M. 2015. Bumblebee Homing: The Fine Structure of Head  
611 Turning Movements. *PLOS ONE* **10**:e0135020. doi:10.1371/journal.pone.0135020
- 612 Brinkworth RS, O’Carroll DC. 2009. Robust models for optic flow coding in natural scenes inspired by insect  
613 biology. *PLoS Comput Biol* **5**:e1000555.
- 614 Burke CJ, Huetteroth W, Oswald D, Perisse E, Krashes MJ, Das G, Gohl D, Silies M, Certel S, Waddell S. 2012.  
615 Layered reward signalling through octopamine and dopamine in Drosophila. *Nature* **492**:433–437.
- 616 Caron SJC, Ruta V, Abbott LF, Axel R. 2013. Random convergence of olfactory inputs in the Drosophila  
617 mushroom body. *Nature* **497**:113–117. doi:10.1038/nature12063
- 618 Cassenaer S, Laurent G. 2012. Conditional modulation of spike-timing-dependent plasticity for olfactory  
619 learning. *Nature* **482**:47–52. doi:10.1038/nature10776
- 620 Cassenaer S, Laurent G. 2007. Hebbian STDP in mushroom bodies facilitates the synchronous flow of  
621 olfactory information in locusts. *Nature* **448**:709–713. doi:10.1038/nature05973
- 622 Chittka L, Niven J. 2009. Are bigger brains better? *Curr Biol CB* **19**:R995–R1008.  
623 doi:10.1016/j.cub.2009.08.023
- 624 Chittka L, Skorupski P. 2017. Active vision: a broader comparative perspective is needed.
- 625 Clark DA, Fitzgerald JE, Ales JM, Gohl DM, Silies MA, Norcia AM, Clandinin TR. 2014. Flies and humans share a  
626 motion estimation strategy that exploits natural scene statistics. *Nat Neurosci* **17**:296–303.  
627 doi:10.1038/nn.3600
- 628 Cognigni P, Felsenberg J, Waddell S. 2018. Do the right thing: neural network mechanisms of memory  
629 formation, expression and update in Drosophila. *Curr Opin Neurobiol* **49**:51–58.  
630 doi:10.1016/j.conb.2017.12.002
- 631 Collett TS, Fry SN, Wehner R. 1993. Sequence learning by honeybees. *J Comp Physiol A* **172**:693–706.  
632 doi:10.1007/BF00195395
- 633 Cope AJ, Vasilaki E, Minors D, Sabo C, Marshall JAR, Barron AB. 2018. Abstract concept learning in a simple  
634 neural network inspired by the insect brain. *PLOS Comput Biol* **14**:e1006435.  
635 doi:10.1371/journal.pcbi.1006435
- 636 Dawkins MS, Woodington A. 2000. Pattern recognition and active vision in chickens. *Nature* **403**:652–655.  
637 doi:10.1038/35001064
- 638 de Croon GCHE, Dupeyroux JG, Fuller SB, Marshall JAR. 2022. Insect-inspired AI for autonomous robots. *Sci*  
639 *Robot* **7**:eabl6334. doi:10.1126/scirobotics.abl6334

- 640 Doussot C, Bertrand OJN, Egelhaaf M. 2021. The Critical Role of Head Movements for Spatial Representation  
641 During Bumblebees Learning Flight. *Front Behav Neurosci* **14**.
- 642 Dyakova O, Lee Y-J, Longden KD, Kiselev VG, Nordström K. 2015. A higher order visual neuron tuned to the  
643 spatial amplitude spectra of natural scenes. *Nat Commun* **6**:8522. doi:10.1038/ncomms9522
- 644 Dyakova O, Müller MM, Egelhaaf M, Nordström K. 2019. Image statistics of the environment surrounding  
645 freely behaving hoverflies. *J Comp Physiol A* **205**:373–385.
- 646 Dyakova O, Nordström K. 2017. Image statistics and their processing in insect vision. *Curr Opin Insect Sci,*  
647 *Neuroscience \* Pheromones* **24**:7–14. doi:10.1016/j.cois.2017.08.002
- 648 Dyer AG, Neumeyer C, Chittka L. 2005. Honeybee (*Apis mellifera*) vision can discriminate between and  
649 recognise images of human faces. *J Exp Biol* **208**:4709–4714.
- 650 Dyer AG, Paulk AC, Reser DH. 2011. Colour processing in complex environments: insights from the visual  
651 system of bees. *Proc R Soc Lond B Biol Sci* **278**:952–959. doi:10.1098/rspb.2010.2412
- 652 Fenk LM, Avritzer SC, Weisman JL, Nair A, Randt LD, Mohren TL, Siwanowicz I, Maimon G. 2022. Muscles that  
653 move the retina augment compound eye vision in *Drosophila*. *Nature* **612**:116–122.  
654 doi:10.1038/s41586-022-05317-5
- 655 Fisher YE, Silies M, Clandinin TR. 2015. Orientation Selectivity Sharpens Motion Detection in *Drosophila*.  
656 *Neuron* **88**:390–402. doi:10.1016/j.neuron.2015.09.033
- 657 Geisler WS. 2008. Visual Perception and the Statistical Properties of Natural Scenes. *Annu Rev Psychol*  
658 **59**:167–192. doi:10.1146/annurev.psych.58.110405.085632
- 659 Gibson JJ. 1979. The ecological approach to visual perception.
- 660 Giger AD, Srinivasan MV. 1996. Pattern recognition in honeybees: chromatic properties of orientation  
661 analysis. *J Comp Physiol A* **178**:763–769. doi:10.1007/BF00225824
- 662 Giurfa M. 2013. Cognition with few neurons: higher-order learning in insects. *Trends Neurosci* **36**:285–294.  
663 doi:10.1016/j.tins.2012.12.011
- 664 Giurfa M, Hammer M, Stach S, Stollhoff N, Müller-Deisig N, Mizyrycki C. 1999. Pattern learning by  
665 honeybees: conditioning procedure and recognition strategy. *Anim Behav* **57**:315–324.  
666 doi:10.1006/anbe.1998.0957
- 667 Giurfa M, Zhang S, Jenett A, Menzel R, Srinivasan MV. 2001. The concepts of ‘sameness’ and ‘difference’ in  
668 an insect. *Nature* **410**:930–933. doi:10.1038/35073582
- 669 Gribakin FG. 1975. Functional morphology of the compound eye of the bee. *Compd Eye Vis Insects* **154**:176.
- 670 Guiraud M, Roper M, Chittka L. 2018. High-Speed Videography Reveals How Honeybees Can Turn a Spatial  
671 Concept Learning Task Into a Simple Discrimination Task by Stereotyped Flight Movements and  
672 Sequential Inspection of Pattern Elements. *Front Psychol* **9**. doi:10.3389/fpsyg.2018.01347
- 673 Hammer M. 1993. An identified neuron mediates the unconditioned stimulus in associative olfactory  
674 learning in honeybees. *Nature* **366**:59–63. doi:10.1038/366059a0
- 675 Hammer M, Menzel R. 1995. Learning and memory in the honeybee. *J Neurosci* **15**:1617–1630.
- 676 Hateren J van, Srinivasan MV, Wait PB. 1990. Pattern recognition in bees: orientation discrimination. *J Comp*  
677 *Physiol A Neuroethol Sens Neural Behav Physiol* **167**:649–654.
- 678 Heisenberg M. 2003. Mushroom body memoir: from maps to models. *Nat Rev Neurosci* **4**:266–275.  
679 doi:10.1038/nrn1074
- 680 Hertel H. 1983. Change of synapse frequency in certain photoreceptors of the honeybee after chromatic  
681 deprivation. *J Comp Physiol* **151**:477–482.
- 682 Hertel H. 1982. The effect of spectral light deprivation on the spectral sensitivity of the honey bee. *J Comp*  
683 *Physiol* **147**:365–369.
- 684 Honegger KS, Campbell RA, Turner GC. 2011. Cellular-resolution population imaging reveals robust sparse  
685 coding in the *Drosophila* mushroom body. *J Neurosci* **31**:11772–11785.
- 686 Hyvärinen A, Hurri J, Hoyer PO. 2009. Natural image statistics: A probabilistic approach to early  
687 computational vision. Springer Science & Business Media.
- 688 James AC, Osorio D. 1996. Characterisation of columnar neurons and visual signal processing in the medulla  
689 of the locust optic lobe by system identification techniques. *J Comp Physiol A* **178**:183–199.  
690 doi:10.1007/BF00188161



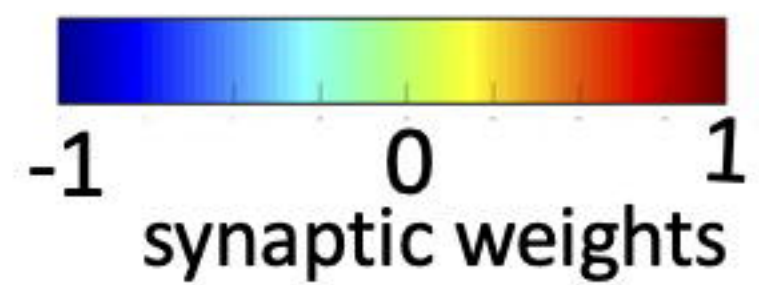
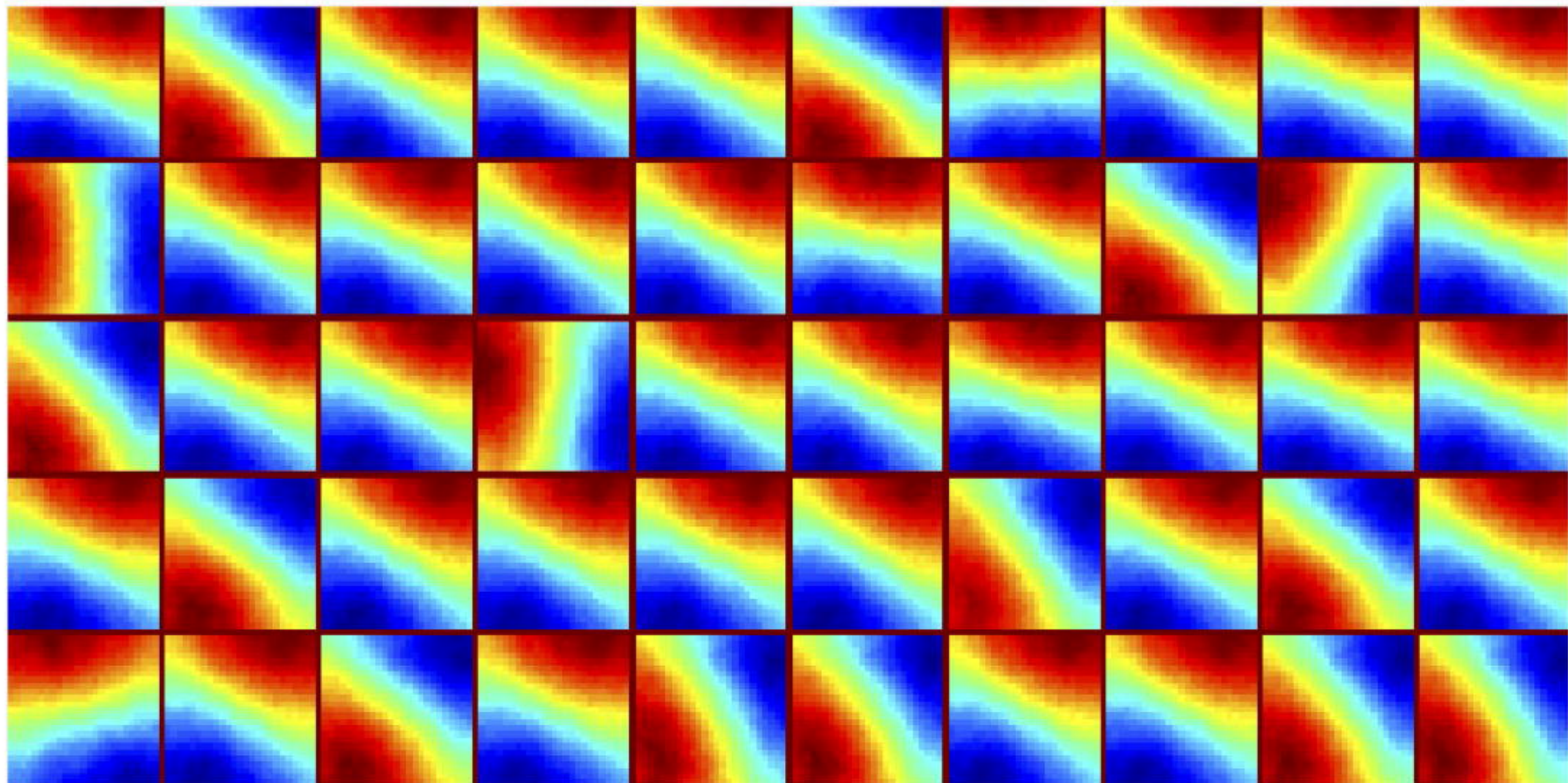
- 691 Juusola M, Dau A, Song Z, Solanki N, Rien D, Jaciuch D, Dongre SA, Blanchard F, Polavieja GG de, Hardie RC,  
692 Takalo J. 2017. Microsaccadic sampling of moving image information provides *Drosophila*  
693 hyperacute vision. *eLife* **6**:e26117. doi:10.7554/eLife.26117
- 694 Juusola M, Song Z. 2017. How a fly photoreceptor samples light information in time. *J Physiol* **595**:5427–  
695 5437.
- 696 Kuang X, Poletti M, Victor JD, Rucci M. 2012. Temporal encoding of spatial information during active visual  
697 fixation. *Curr Biol CB* **22**:510–514. doi:10.1016/j.cub.2012.01.050
- 698 Land MF. 1999. Motion and vision: why animals move their eyes. *J Comp Physiol [A]* **185**:341–352.
- 699 Land MF. 1973. Head Movement of Flies during Visually Guided Flight. *Nature* **243**:299–300.  
700 doi:10.1038/243299a0
- 701 Land MF, Nilsson D-E. 2012. Animal eyes. Oxford University Press.
- 702 Langridge KV, Wilke C, Riabinina O, Vorobyev M, de Ibarra NH. 2018. Approach direction prior to landing  
703 explains patterns of colour learning. *bioRxiv* 381210.
- 704 Langridge KV, Wilke C, Riabinina O, Vorobyev M, Hempel de Ibarra N. 2021. Approach Direction Prior to  
705 Landing Explains Patterns of Colour Learning in Bees. *Front Physiol* **12**.
- 706 Lehrer M, Collett TS. 1994. Approaching and departing bees learn different cues to the distance of a  
707 landmark. *J Comp Physiol A* **175**:171–177. doi:10.1007/BF00215113
- 708 Locatelli FF, Fernandez PC, Villareal F, Muezzinoglu K, Huerta R, Galizia CG, Smith BH. 2013. Nonassociative  
709 plasticity alters competitive interactions among mixture components in early olfactory processing.  
710 *Eur J Neurosci* **37**:63–79. doi:10.1111/ejn.12021
- 711 MaBouDi H, Barron AB, Li S, Honkanen M, Loukola OJ, Peng F, Li W, Marshall JAR, Cope A, Vasilaki E, Solvi C.  
712 2021a. Non-numerical strategies used by bees to solve numerical cognition tasks. *Proc R Soc B Biol*  
713 *Sci* **288**:20202711. doi:10.1098/rspb.2020.2711
- 714 MaBouDi H, Dona H, Gatto E, Loukola OJ, Buckley E, Onoufriou PD, Skorupski P, Chittka L. 2020a.  
715 Bumblebees use sequential scanning of countable items in visual patterns to solve numerosity tasks.  
716 *Integr Comp Biol*.
- 717 MaBouDi H, Marshall JA, Dearden N, Barron AB. 2023. How honey bees make fast and accurate decisions.  
718 *bioRxiv* 2023–01.
- 719 MaBouDi H, Marshall JAR, Barron AB. 2020b. Honeybees solve a multi-comparison ranking task by  
720 probability matching. *Proc R Soc B Biol Sci* **287**:20201525. doi:10.1098/rspb.2020.1525
- 721 MaBouDi H, Roper M, Guiraud M, Marshall JAR, Chittka L. 2021b. Automated video tracking and flight  
722 analysis show how bumblebees solve a pattern discrimination task using active vision. *bioRxiv*  
723 2021.03.09.434580. doi:10.1101/2021.03.09.434580
- 724 MaBouDi H, Shimazaki H, Giurfa M, Chittka L. 2017. Olfactory learning without the mushroom bodies:  
725 Spiking neural network models of the honeybee lateral antennal lobe tract reveal its capacities in  
726 odour memory tasks of varied complexities. *PLoS Comput Biol* **13**:e1005551.
- 727 MaBouDi H, Solvi C, Chittka L. 2020c. Bumblebees learn a relational rule but switch to a win-stay/lose-switch  
728 heuristic after extensive training. *bioRxiv* 2020.05.08.085142. doi:10.1101/2020.05.08.085142
- 729 Markram H, Lübke J, Frotscher M, Sakmann B. 1997. Regulation of synaptic efficacy by coincidence of  
730 postsynaptic APs and EPSPs. *Science* **275**:213–215.
- 731 Matthews T, Osorio D, Cavallaro A, Chittka L. 2018. The importance of spatial visual scene parameters in  
732 predicting optimal cone sensitivities in routinely trichromatic frugivorous Old-World primates. *Front*  
733 *Comput Neurosci* **12**:15.
- 734 Meeks JP, Holy TE. 2008. Pavlov's moth: olfactory learning and spike timing-dependent plasticity. *Nat*  
735 *Neurosci* **11**:1126–1127.
- 736 Menzel R. 2012. The honeybee as a model for understanding the basis of cognition. *Nat Rev Neurosci*  
737 **13**:758–768. doi:10.1038/nrn3357
- 738 Menzel R, Blakers M. 1976. Colour receptors in the bee eye — Morphology and spectral sensitivity. *J Comp*  
739 *Physiol* **108**:11–13. doi:10.1007/BF00625437
- 740 Menzel R, Giurfa M. 2006. Dimensions of cognition in an insect, the honeybee. *Behav Cogn Neurosci Rev*  
741 **5**:24–40. doi:10.1177/1534582306289522

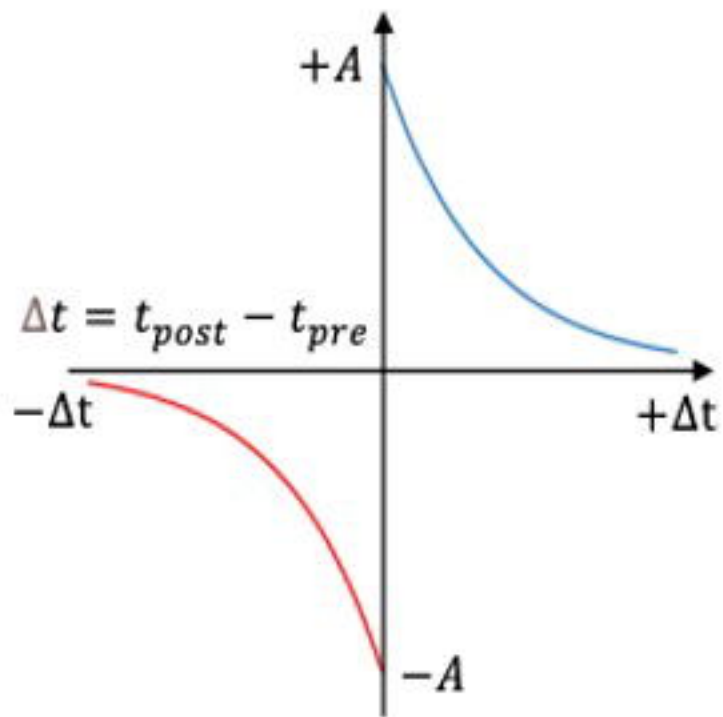
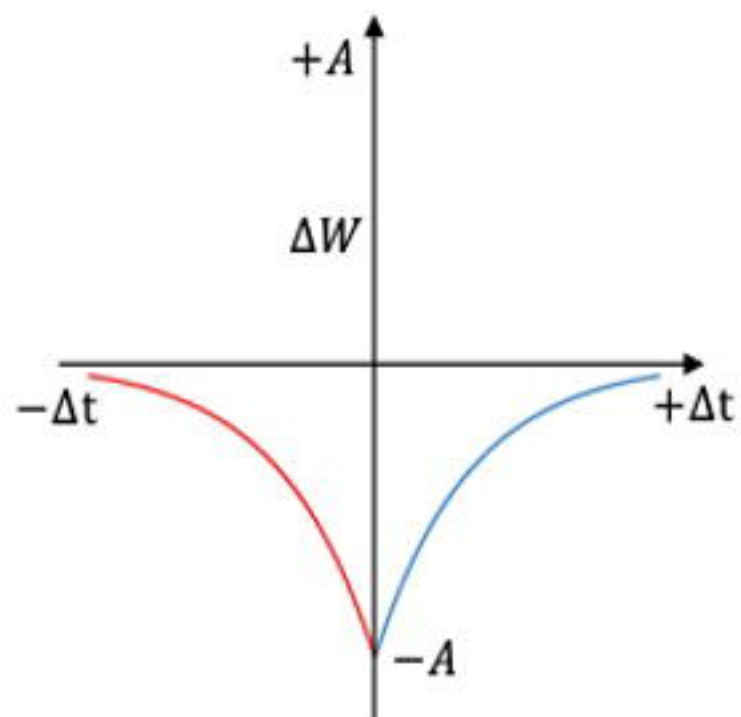


- 742 Nityananda V, Skorupski P, Chittka L. 2014. Can bees see at a glance? *J Exp Biol* **217**:1933–1939.  
743 doi:10.1242/jeb.101394
- 744 Odenthal L, Doussot C, Meyer S, Bertrand OJN. 2021. Analysing Head-Thorax Choreography During Free-  
745 Flights in Bumblebees. *Front Behav Neurosci* **14**. doi:10.3389/fnbeh.2020.610029
- 746 Oja E. 1982. Simplified neuron model as a principal component analyzer. *J Math Biol* **15**:267–273.
- 747 Okada R, Rybak J, Manz G, Menzel R. 2007. Learning-Related Plasticity in PE1 and Other Mushroom Body-  
748 Extrinsic Neurons in the Honeybee Brain. *J Neurosci* **27**:11736–11747. doi:10.1523/JNEUROSCI.2216-  
749 07.2007
- 750 Olshausen BA. 2003. Principles of image representation in visual cortex. *Vis Neurosci* **2**:1603–1615.
- 751 Paulk AC, Dacks AM, Phillips-Portillo J, Fellous J-M, Gronenberg W. 2009. Visual Processing in the Central Bee  
752 Brain. *J Neurosci* **29**:9987–9999. doi:10.1523/JNEUROSCI.1325-09.2009
- 753 Paulk AC, Gronenberg W. 2008. Higher order visual input to the mushroom bodies in the bee, *Bombus*  
754 *impatiens*. *Arthropod Struct Dev* **37**:443–458. doi:10.1016/j.asd.2008.03.002
- 755 Paulk AC, Phillips-Portillo J, Dacks AM, Fellous J-M, Gronenberg W. 2008. The Processing of Color, Motion,  
756 and Stimulus Timing Are Anatomically Segregated in the Bumblebee Brain. *J Neurosci* **28**:6319–6332.  
757 doi:10.1523/JNEUROSCI.1196-08.2008
- 758 Perry CJ, Barron AB. 2013. Neural mechanisms of reward in insects. *Annu Rev Entomol* **58**:543–562.
- 759 Roper M, Fernando C, Chittka L. 2017. Insect Bio-inspired Neural Network Provides New Evidence on How  
760 Simple Feature Detectors Can Enable Complex Visual Generalization and Stimulus Location  
761 Invariance in the Miniature Brain of Honeybees. *PLOS Comput Biol* **13**:e1005333.  
762 doi:10.1371/journal.pcbi.1005333
- 763 Rucci M, Iovin R, Poletti M, Santini F. 2007. Miniature eye movements enhance fine spatial detail. *Nature*  
764 **447**:852–855.
- 765 Rucci M, Victor JD. 2015. The unsteady eye: an information-processing stage, not a bug. *Trends Neurosci*  
766 **38**:195–206. doi:10.1016/j.tins.2015.01.005
- 767 Ruderman DL. 1994. The statistics of natural images. *Netw Comput Neural Syst* **5**:517.
- 768 Schwaerzel M, Monastirioti M, Scholz H, Friggi-Grelin F, Birman S, Heisenberg M. 2003. Dopamine and  
769 octopamine differentiate between aversive and appetitive olfactory memories in *Drosophila*. *J*  
770 *Neurosci Off J Soc Neurosci* **23**:10495–10502.
- 771 Schwegmann A, Lindemann JP, Egelhaaf M. 2014. Temporal statistics of natural image sequences generated  
772 by movements with insect flight characteristics. *PloS One* **9**:e110386.  
773 doi:10.1371/journal.pone.0110386
- 774 Seelig JD, Jayaraman V. 2013. Feature detection and orientation tuning in the *Drosophila* central complex.  
775 *Nature* **503**:262–266. doi:10.1038/nature12601
- 776 Serbe E, Meier M, Leonhardt A, Borst A. 2016. Comprehensive characterization of the major presynaptic  
777 elements to the *Drosophila* OFF motion detector. *Neuron* **89**:829–841.
- 778 Simoncelli EP, Olshausen BA. 2001. Natural image statistics and neural representation. *Annu Rev Neurosci*  
779 **24**:1193–1216.
- 780 Skorupski P, Döring TF, Chittka L. 2007. Photoreceptor spectral sensitivity in island and mainland populations  
781 of the bumblebee, *Bombus terrestris*. *J Comp Physiol A* **193**:485–494.
- 782 Song S, Miller KD, Abbott LF. 2000. Competitive Hebbian learning through spike-timing-dependent synaptic  
783 plasticity. *Nat Neurosci* **3**:919–926.
- 784 Song Z, Juusola M. 2014. Refractory Sampling Links Efficiency and Costs of Sensory Encoding to Stimulus  
785 Statistics. *J Neurosci* **34**:7216–7237. doi:10.1523/JNEUROSCI.4463-13.2014
- 786 Spaethe J, Chittka L. 2003. Interindividual variation of eye optics and single object resolution in bumblebees.  
787 *J Exp Biol* **206**:3447–3453.
- 788 Spaethe J, Tautz J, Chittka L. 2001. Visual constraints in foraging bumblebees: flower size and color affect  
789 search time and flight behavior. *Proc Natl Acad Sci* **98**:3898–3903.
- 790 Srinivasan MV. 2010. Honey bees as a model for vision, perception, and cognition. *Annu Rev Entomol*  
791 **55**:267–284.
- 792 Srinivasan MV. 1994. Pattern recognition in the honeybee: recent progress. *J Insect Physiol* **40**:183–194.

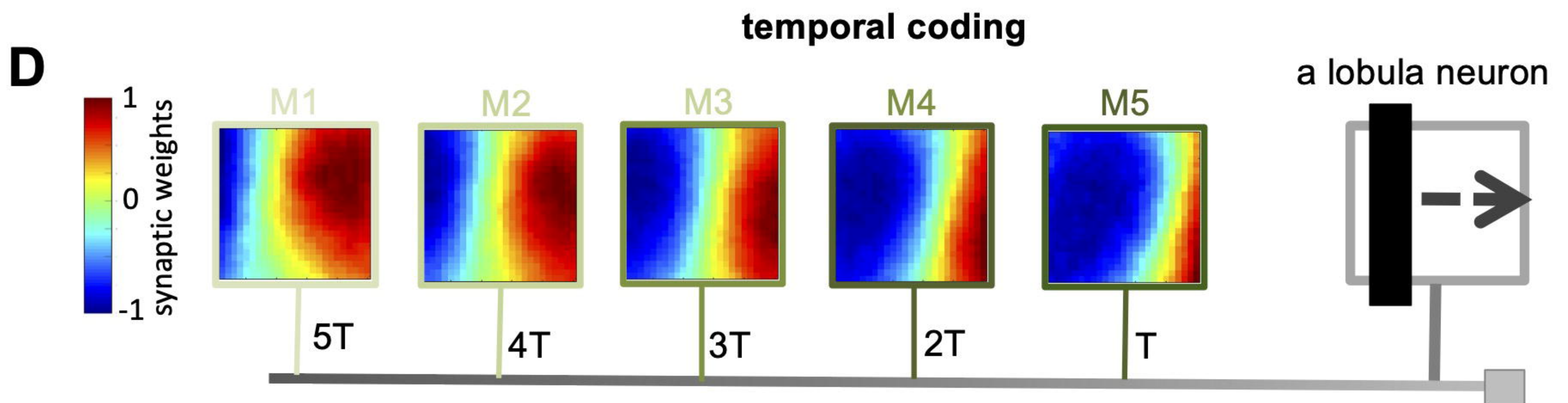
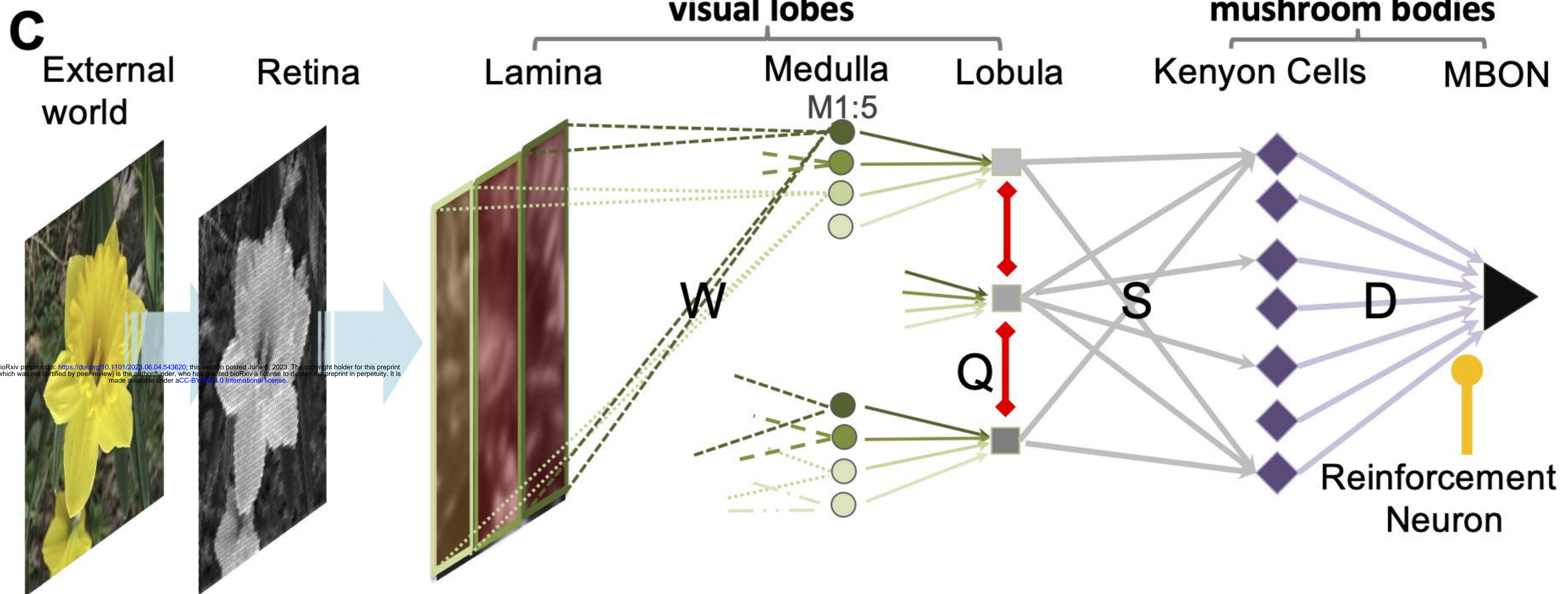
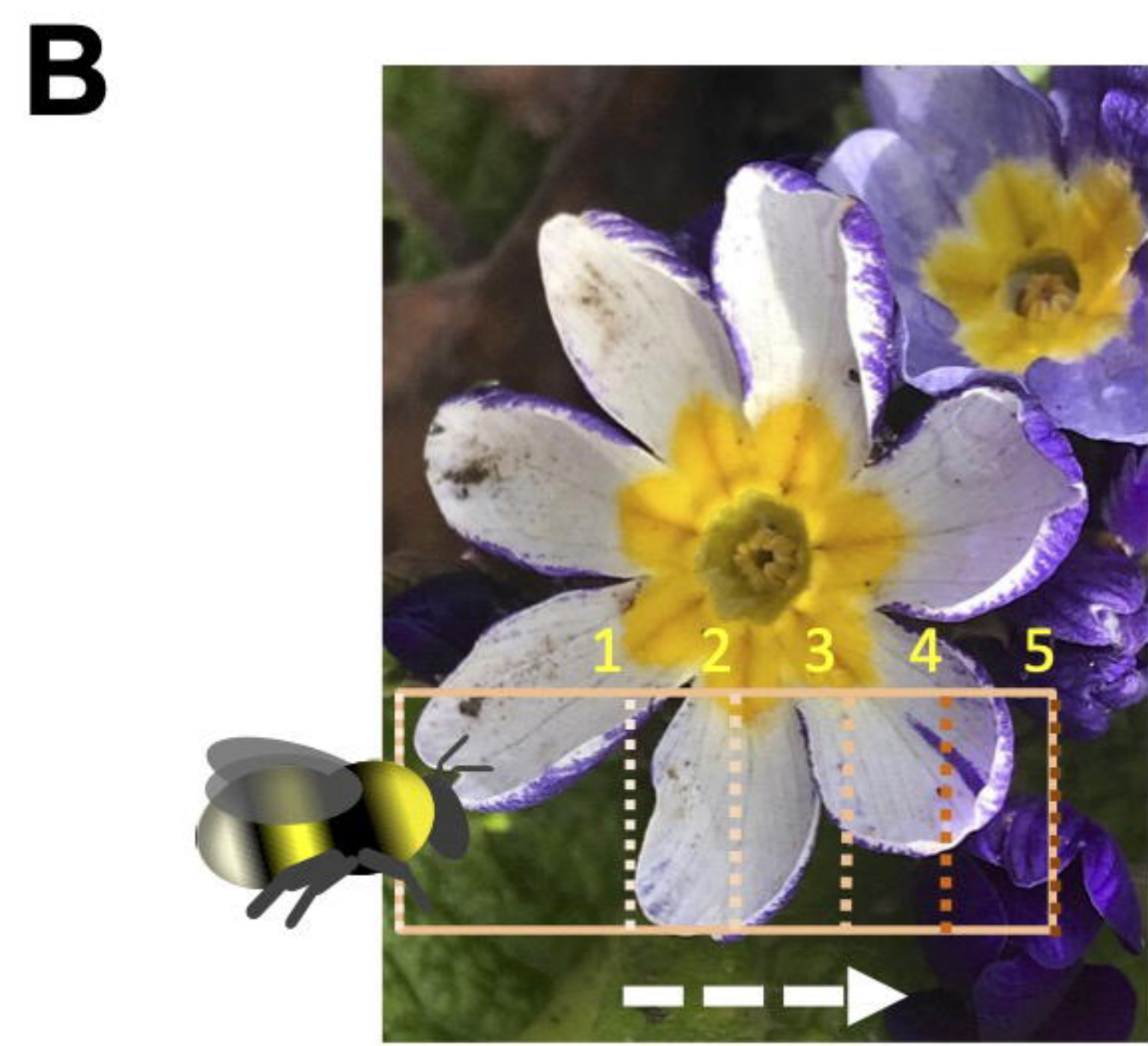
- 793 Srinivasan MV, Lehrer M. 1988. Spatial acuity of honeybee vision and its spectral properties. *J Comp Physiol*  
794 *A* **162**:159–172. doi:10.1007/BF00606081
- 795 Stach S, Benard J, Giurfa M. 2004. Local-feature assembling in visual pattern recognition and generalization  
796 in honeybees. *Nature* **429**:758–761. doi:10.1038/nature02594
- 797 Stach S, Giurfa M. 2005. The influence of training length on generalization of visual feature assemblies in  
798 honeybees. *Behav Brain Res* **161**:8–17. doi:10.1016/j.bbr.2005.02.008
- 799 Streinzer M, Brockmann A, Nagaraja N, Spaethe J. 2013. Sex and Caste-Specific Variation in Compound Eye  
800 Morphology of Five Honeybee Species. *PLOS ONE* **8**:e57702. doi:10.1371/journal.pone.0057702
- 801 Strube-Bloss MF, Nawrot MP, Menzel R. 2011. Mushroom Body Output Neurons Encode Odor–Reward  
802 Associations. *J Neurosci* **31**:3129–3140. doi:10.1523/JNEUROSCI.2583-10.2011
- 803 Szyszka P, Ditzen M, Galkin A, Galizia CG, Menzel R. 2005. Sparsening and temporal sharpening of olfactory  
804 representations in the honeybee mushroom bodies. *J Neurophysiol* **94**:3303–3313.  
805 doi:10.1152/jn.00397.2005
- 806 Taylor GJ, Tichit P, Schmidt MD, Bodey AJ, Rau C, Baird E. 2019. Bumblebee visual allometry results in locally  
807 improved resolution and globally improved sensitivity. *Elife* **8**:e40613.
- 808 Turner CH. 1911. Experiments on Pattern-Vision of the Honey Bee. *Biol Bull* **21**:249–264.  
809 doi:10.2307/1536017
- 810 Van Hateren JH. 1997. Processing of natural time series of intensities by the visual system of the blowfly.  
811 *Vision Res* **37**:3407–3416.
- 812 van Hateren JH. 1992. Theoretical predictions of spatiotemporal receptive fields of fly LMCs, and  
813 experimental validation. *J Comp Physiol A* **171**:157–170.
- 814 Vasas V, Peng F, MaBouDi H, Chittka L. 2019. Randomly weighted receptor inputs can explain the large  
815 diversity of colour-coding neurons in the bee visual system. *Sci Rep* **9**:1–13.
- 816 Vetter RS, Visscher PK. n.d. Influence of Age on Antennal Response of Male Honey Bees, *Apis mellifera*, to  
817 Queen Mandibular Pheromone and Alarm Pheromone Component. *J Chem Ecol* **23**:1867–1880.  
818 doi:10.1023/B:JOEC.0000006456.90528.94
- 819 Vogels TP, Sprekeler H, Zenke F, Clopath C, Gerstner W. 2011. Inhibitory Plasticity Balances Excitation and  
820 Inhibition in Sensory Pathways and Memory Networks. *Science* **334**:1569–1573.  
821 doi:10.1126/science.1211095
- 822 Von Frisch K. 1914. Der farbensinn und formensinn der biene. Рипол Классик.
- 823 Webb B. 2020. Robots with insect brains. *Science* **368**:244–245.
- 824 Wehner R. 1967. Pattern recognition in bees. *Nature* **215**:1244–1248.
- 825 Werner A, Stürzl W, Zanker J. 2016. Object Recognition in Flight: How Do Bees Distinguish between 3D  
826 Shapes? *PLoS ONE* **11**:e0147106. doi:10.1371/journal.pone.0147106
- 827 Yang E-C, Maddess T. 1997. Orientation-sensitive Neurons in the Brain of the Honey Bee (*Apis mellifera*). *J*  
828 *Insect Physiol* **43**:329–336. doi:10.1016/S0022-1910(96)00111-4
- 829 Yarbus AL. 2013. Eye movements and vision. Springer.
- 830 Zhang LI, Tao HW, Holt CE, Harris WA, Poo M. 1998. A critical window for cooperation and competition  
831 among developing retinotectal synapses. *Nature* **395**:37–44.
- 832 Zhang SW, Horridge GA. 1992. Pattern recognition in bees: size of regions in spatial layout. *Philos Trans R Soc*  
833 *Lond B Biol Sci* **337**:65–71.
- 834 Zimmermann MJY, Nevala NE, Yoshimatsu T, Osorio D, Nilsson D-E, Berens P, Baden T. 2018. Zebrafish  
835 Differentially Process Color across Visual Space to Match Natural Scenes. *Curr Biol* **0**.  
836 doi:10.1016/j.cub.2018.04.075  
837



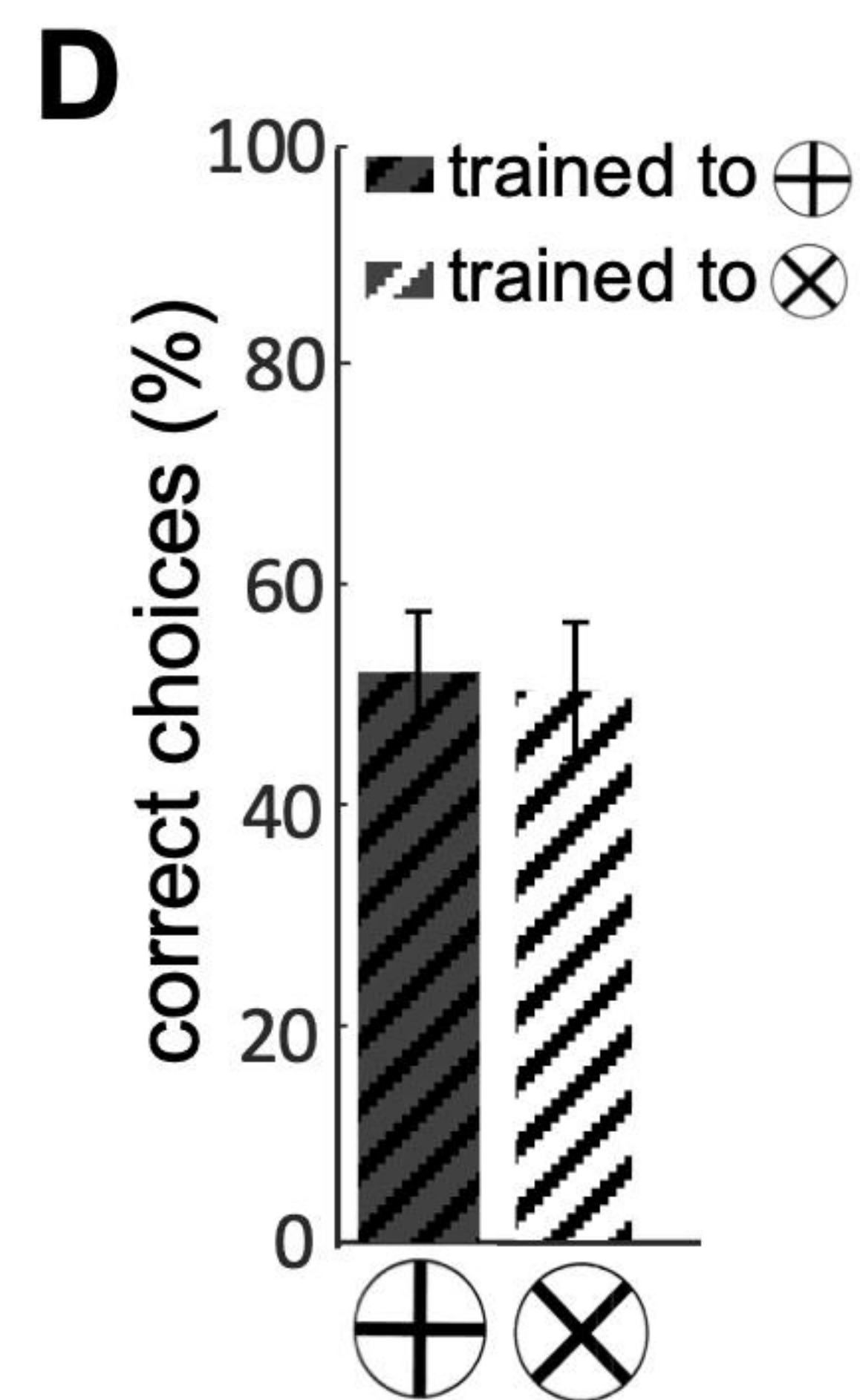
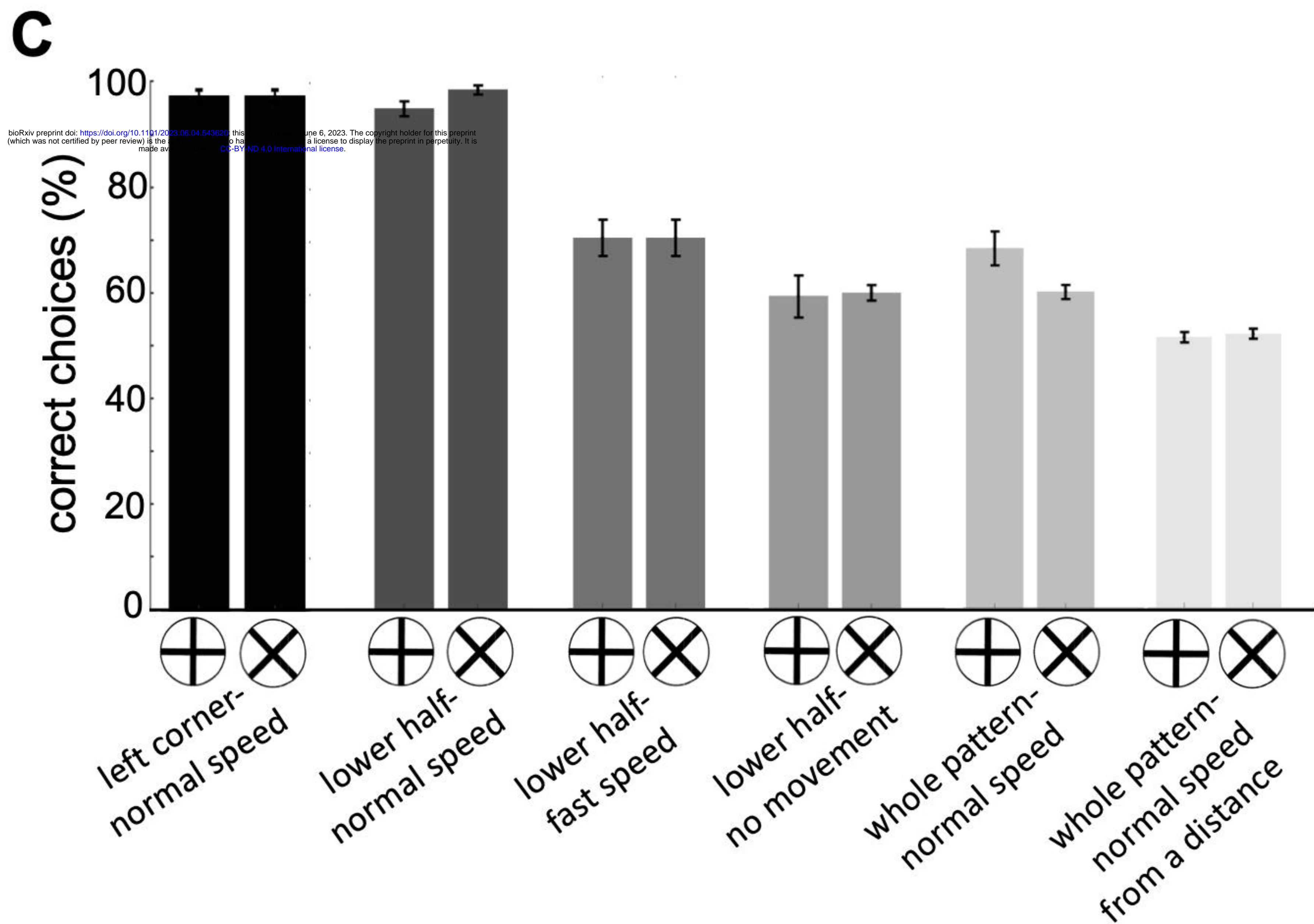
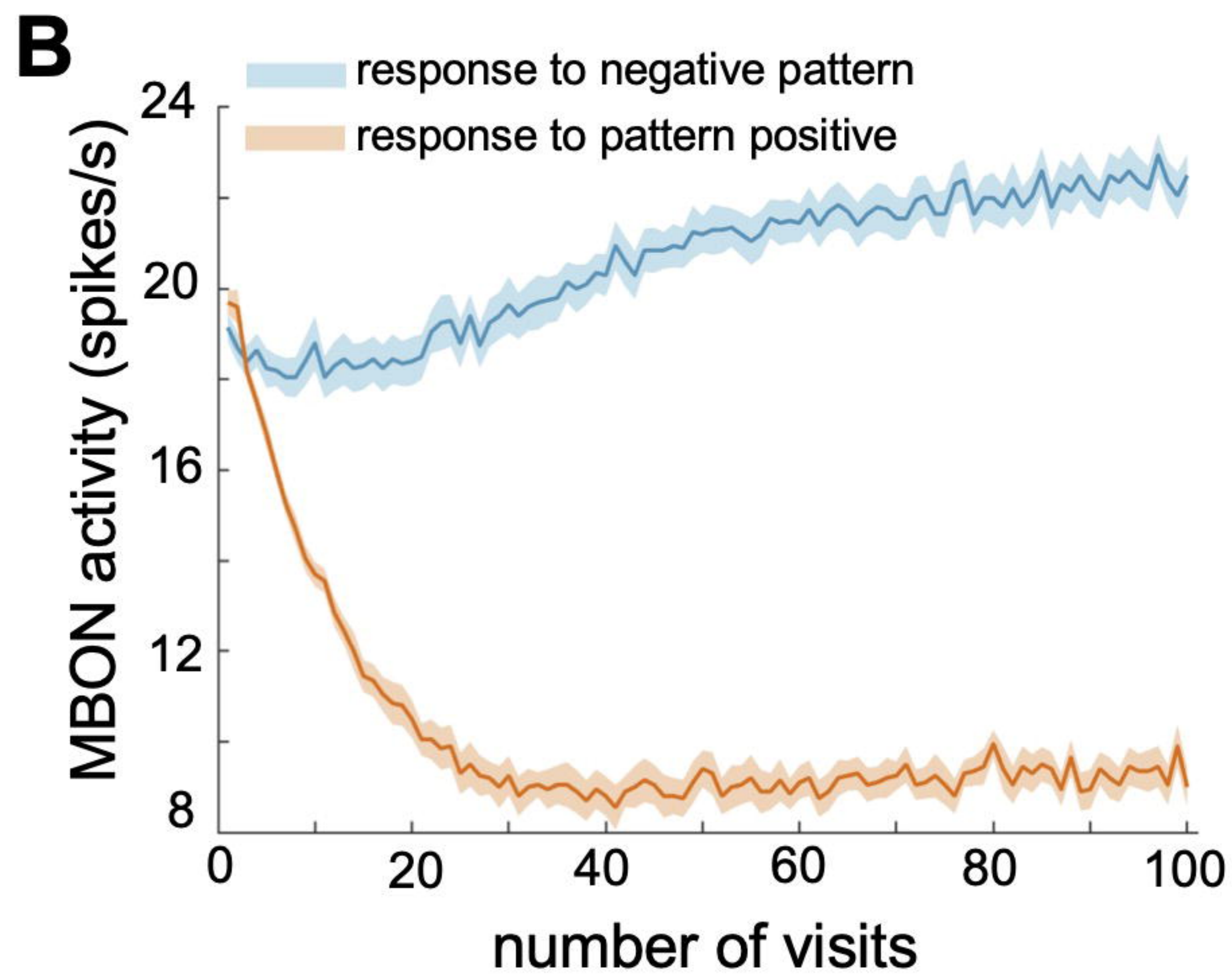
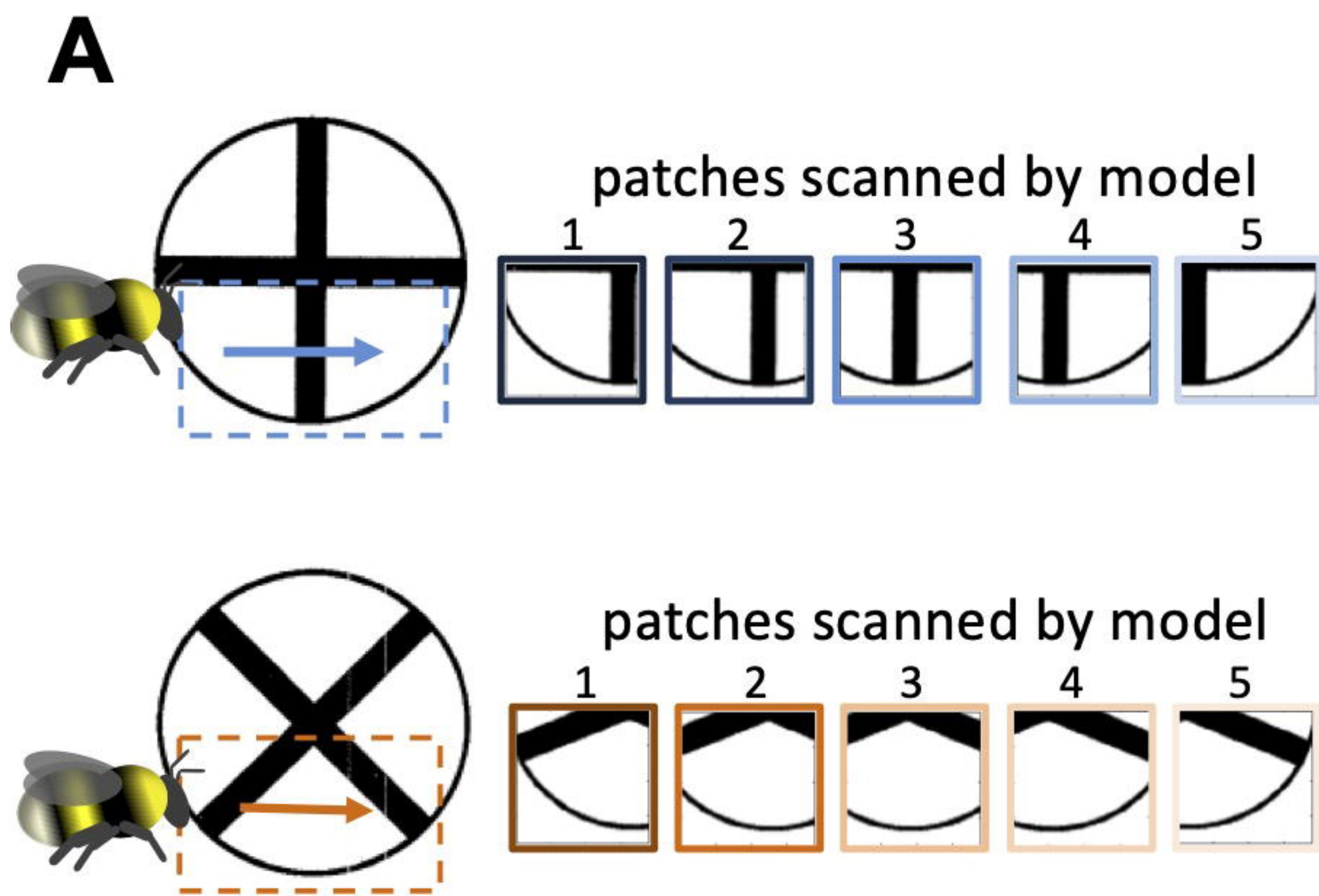


**A****B**

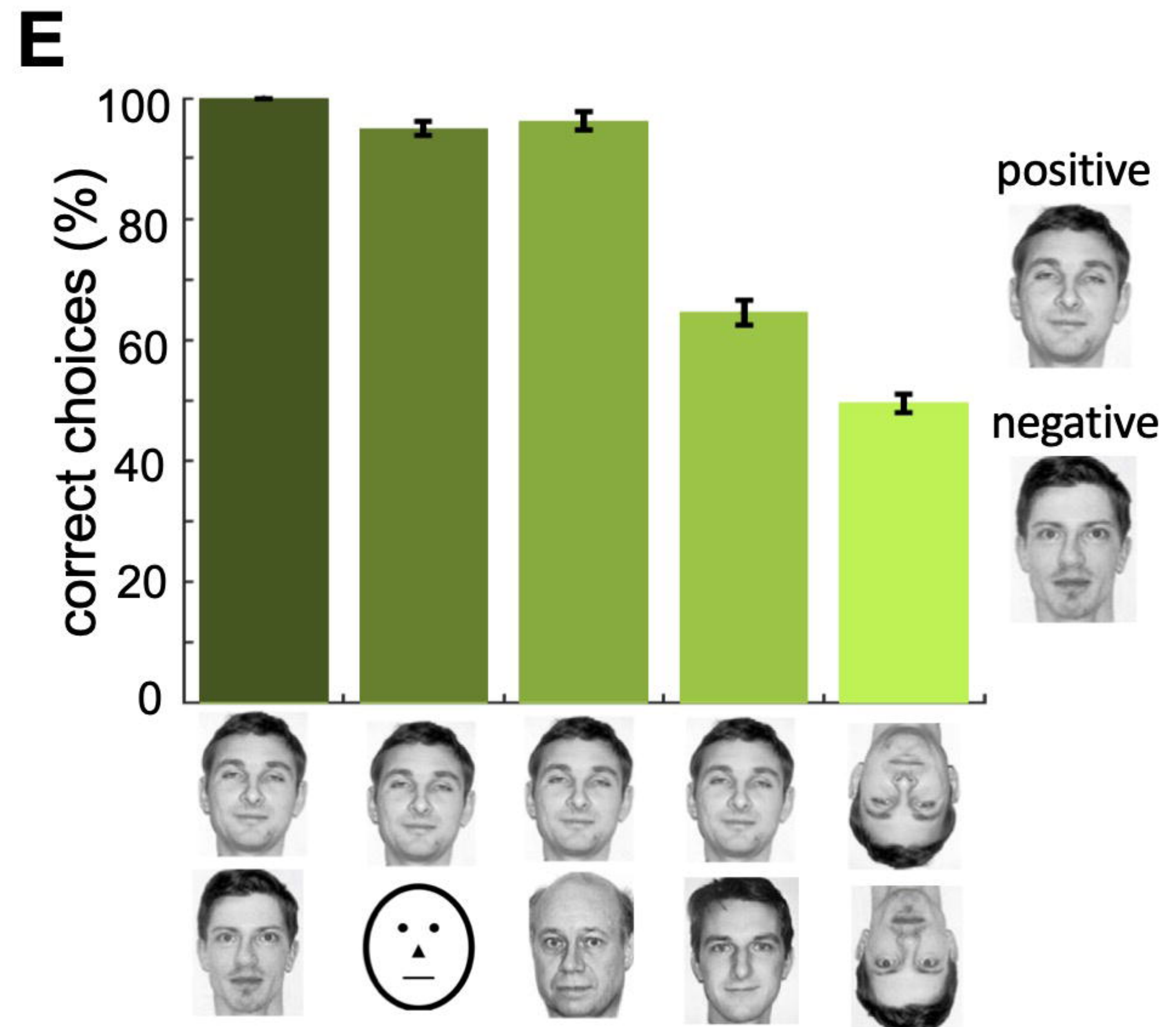
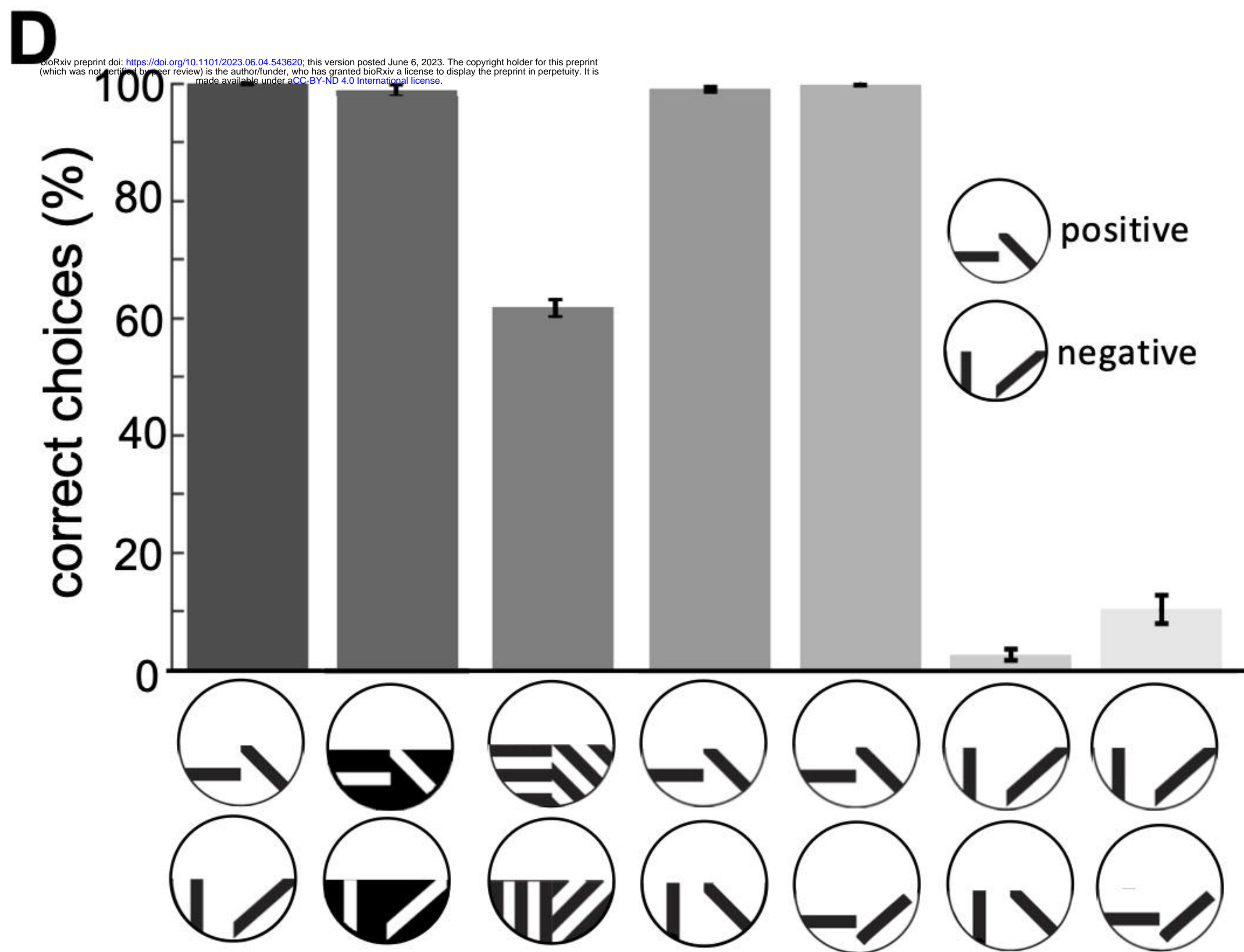
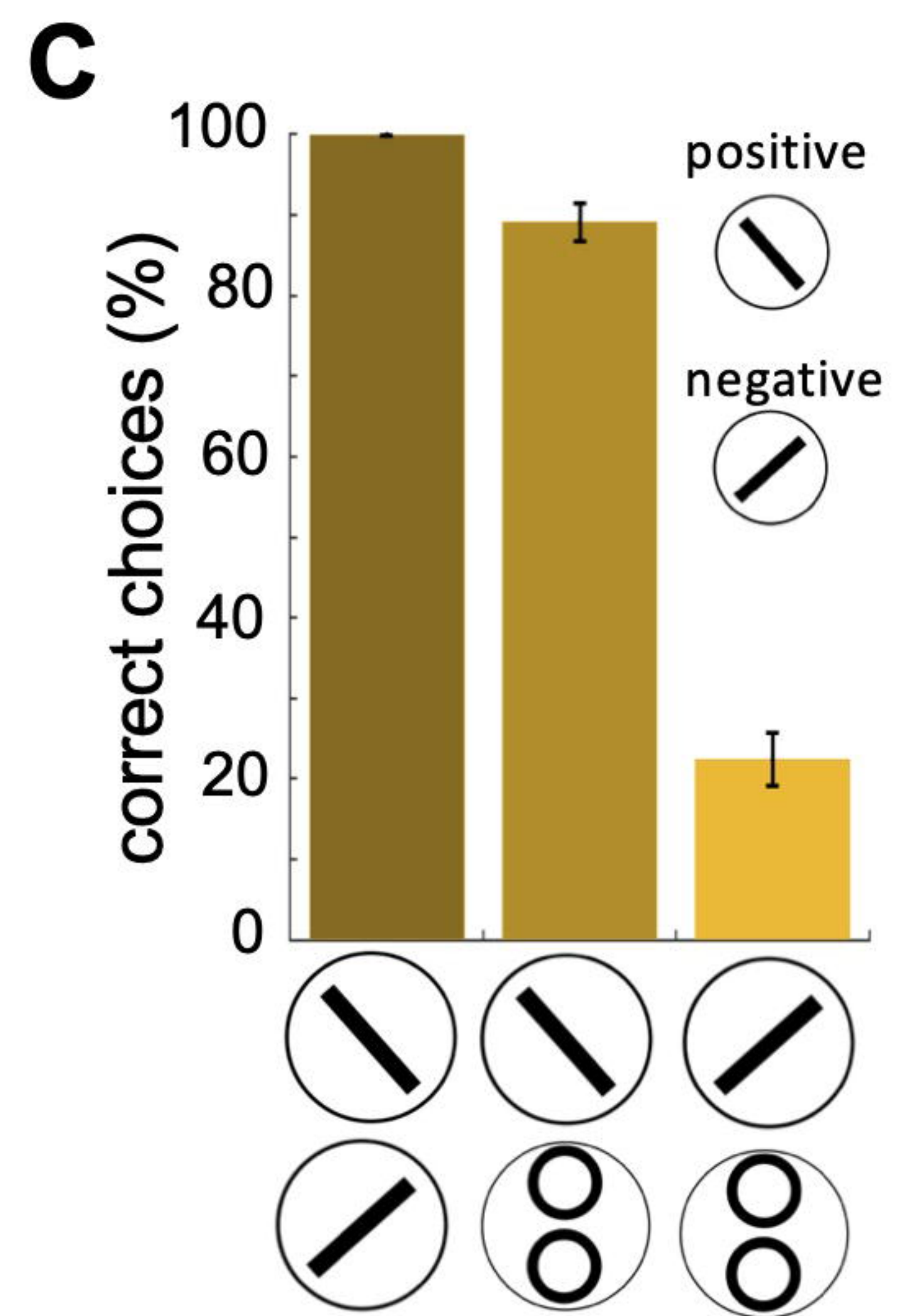
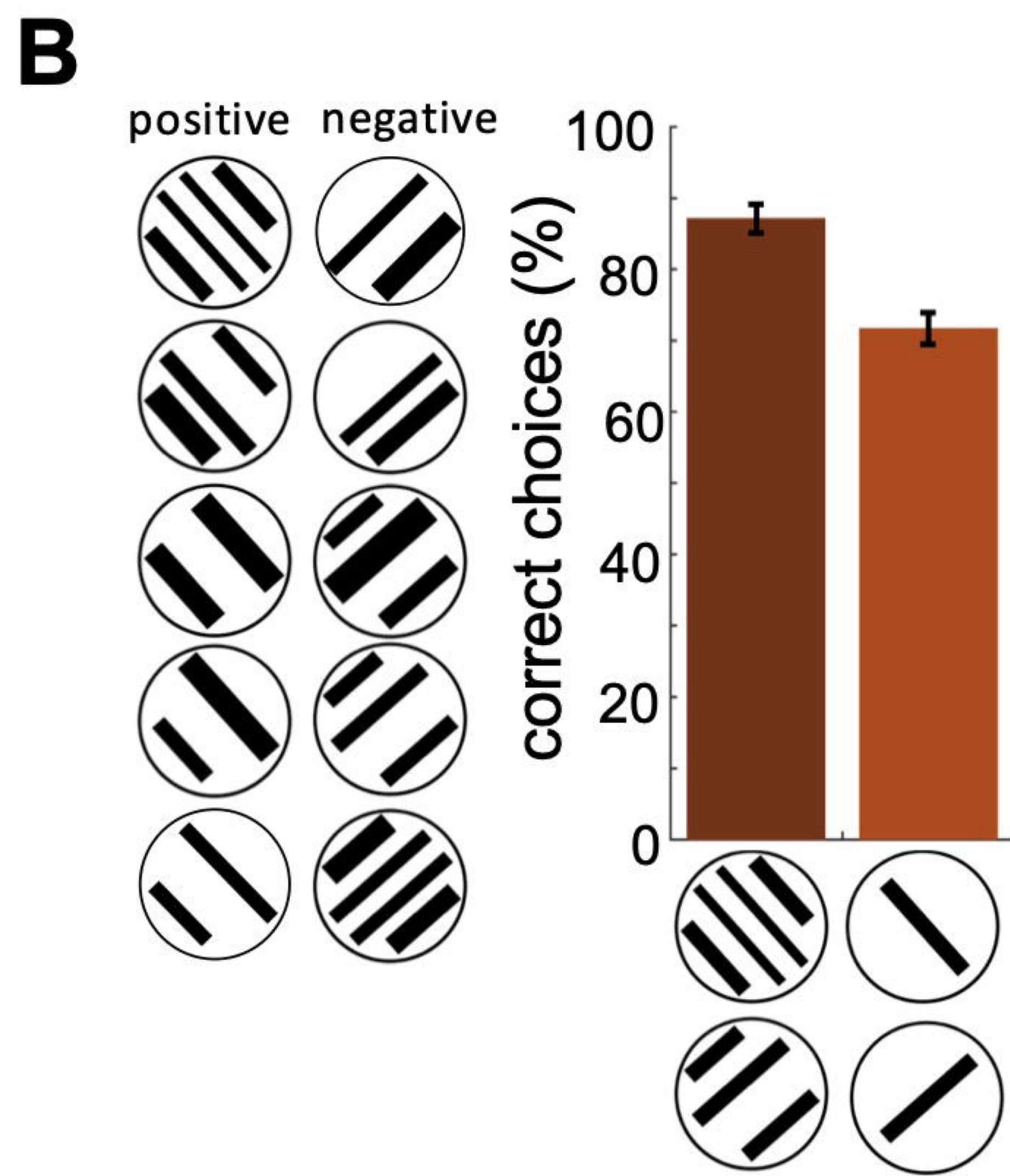
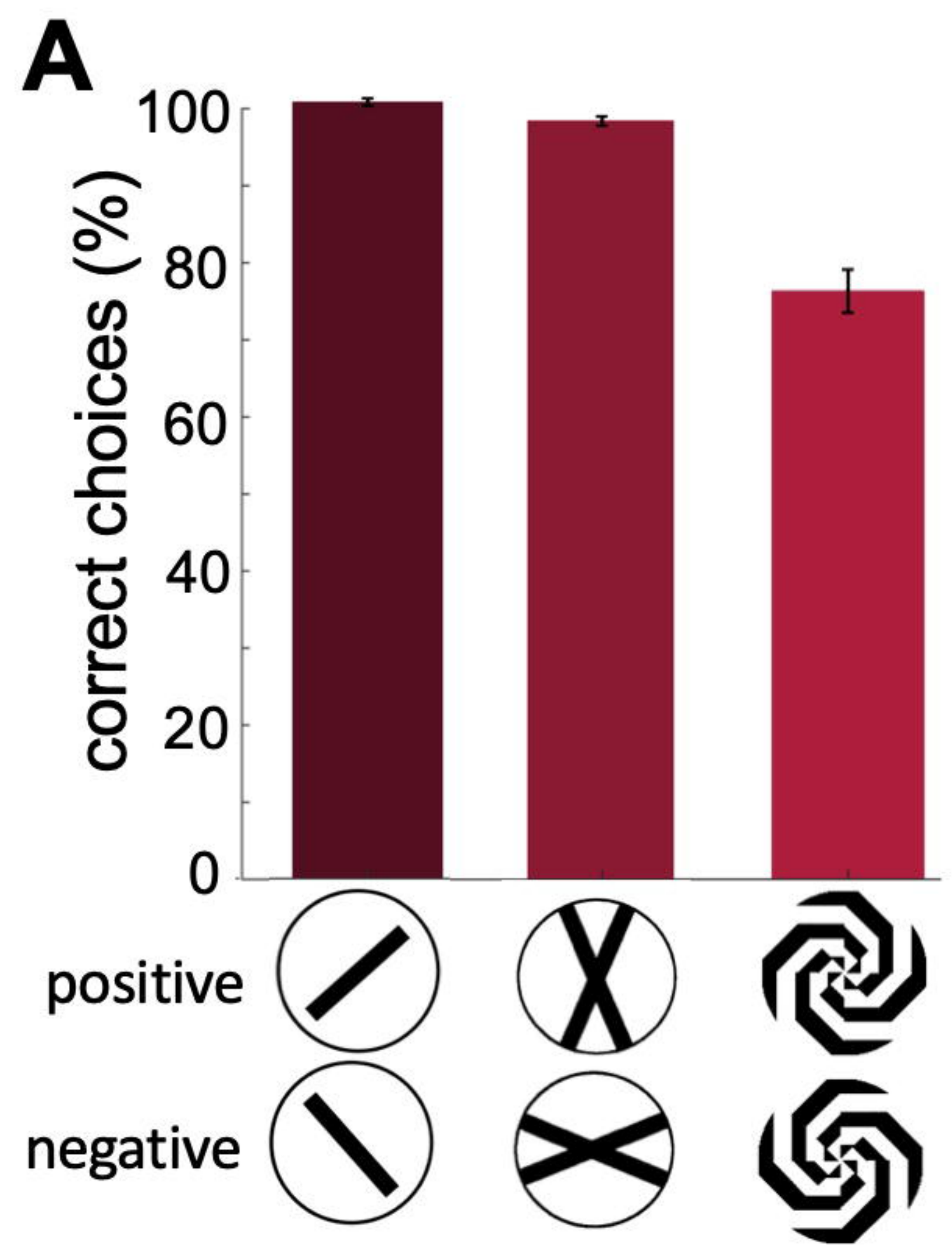




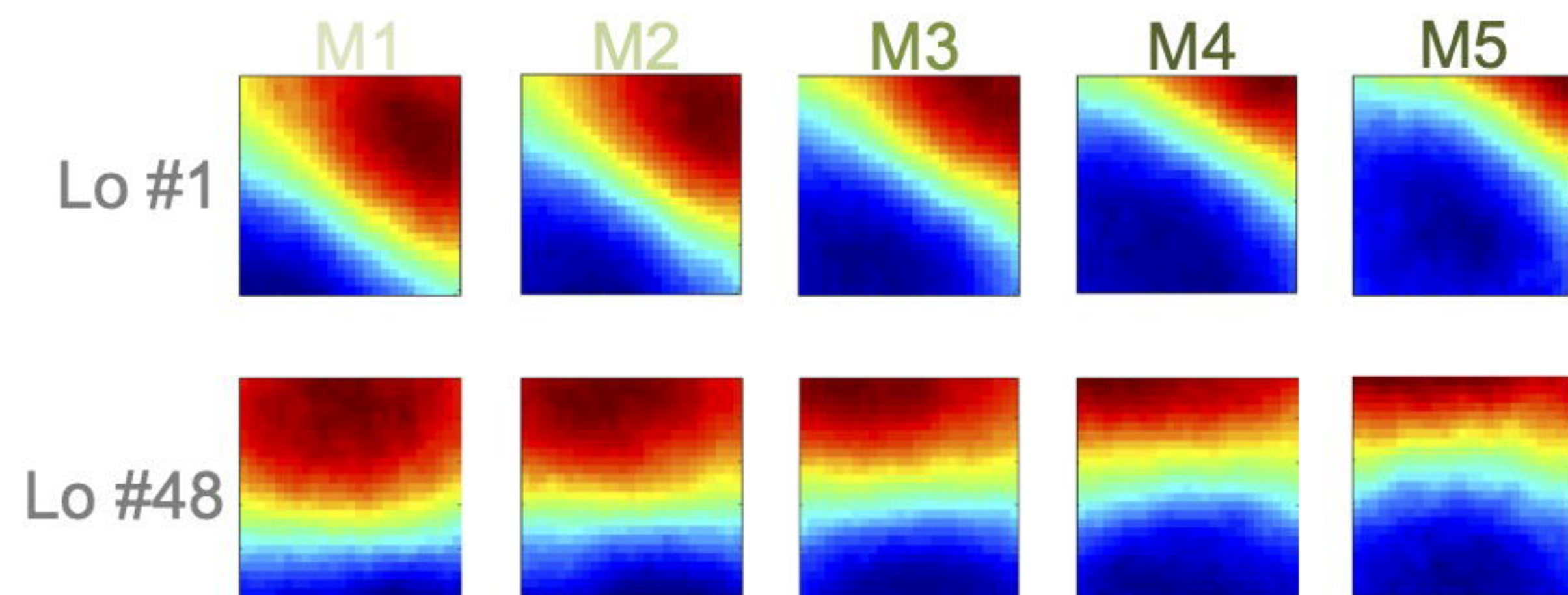
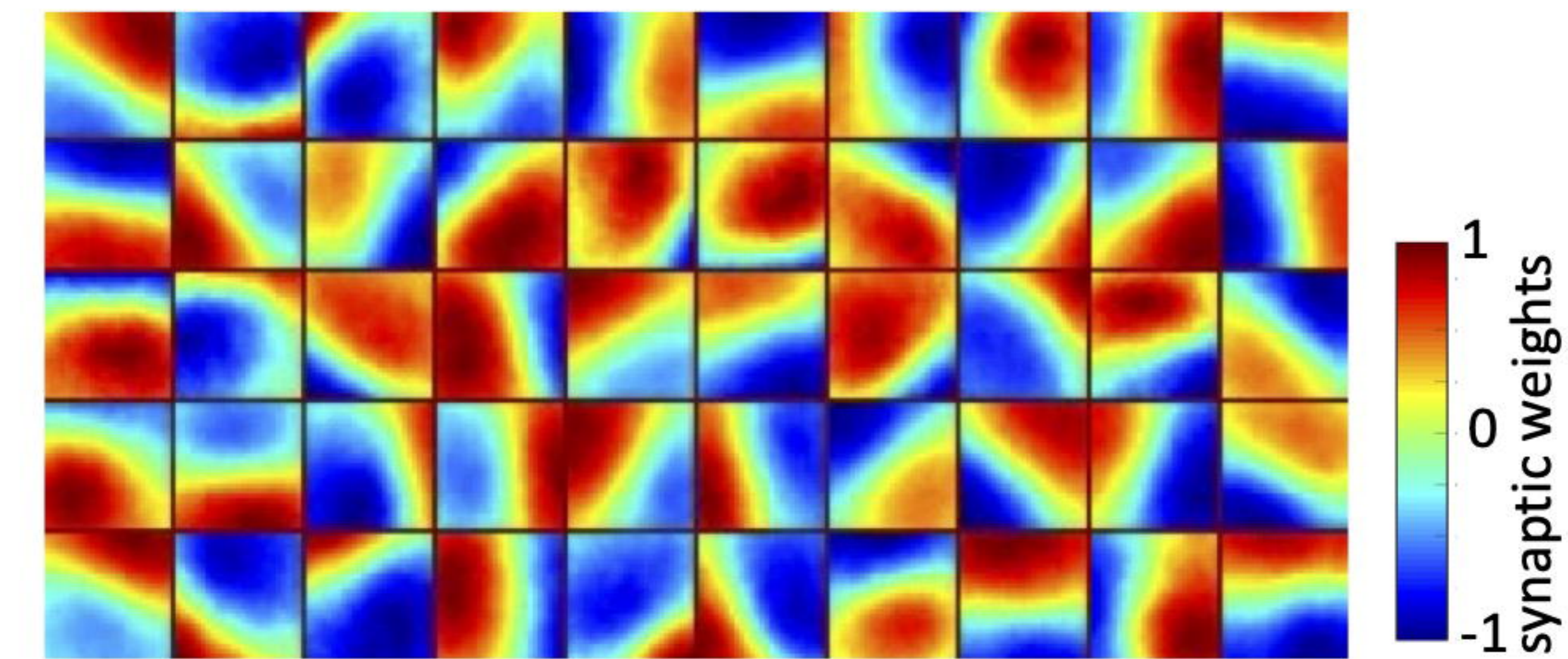
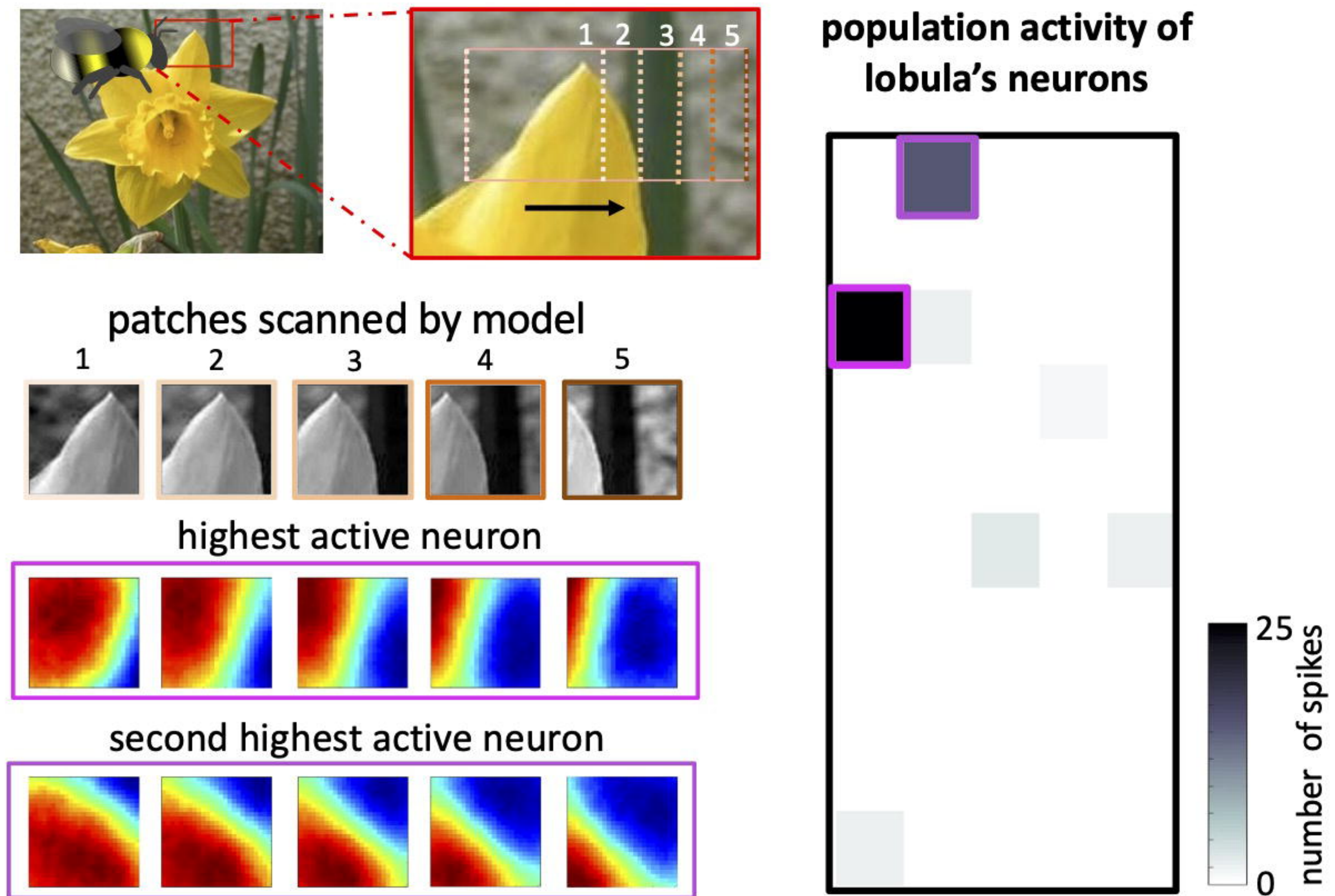
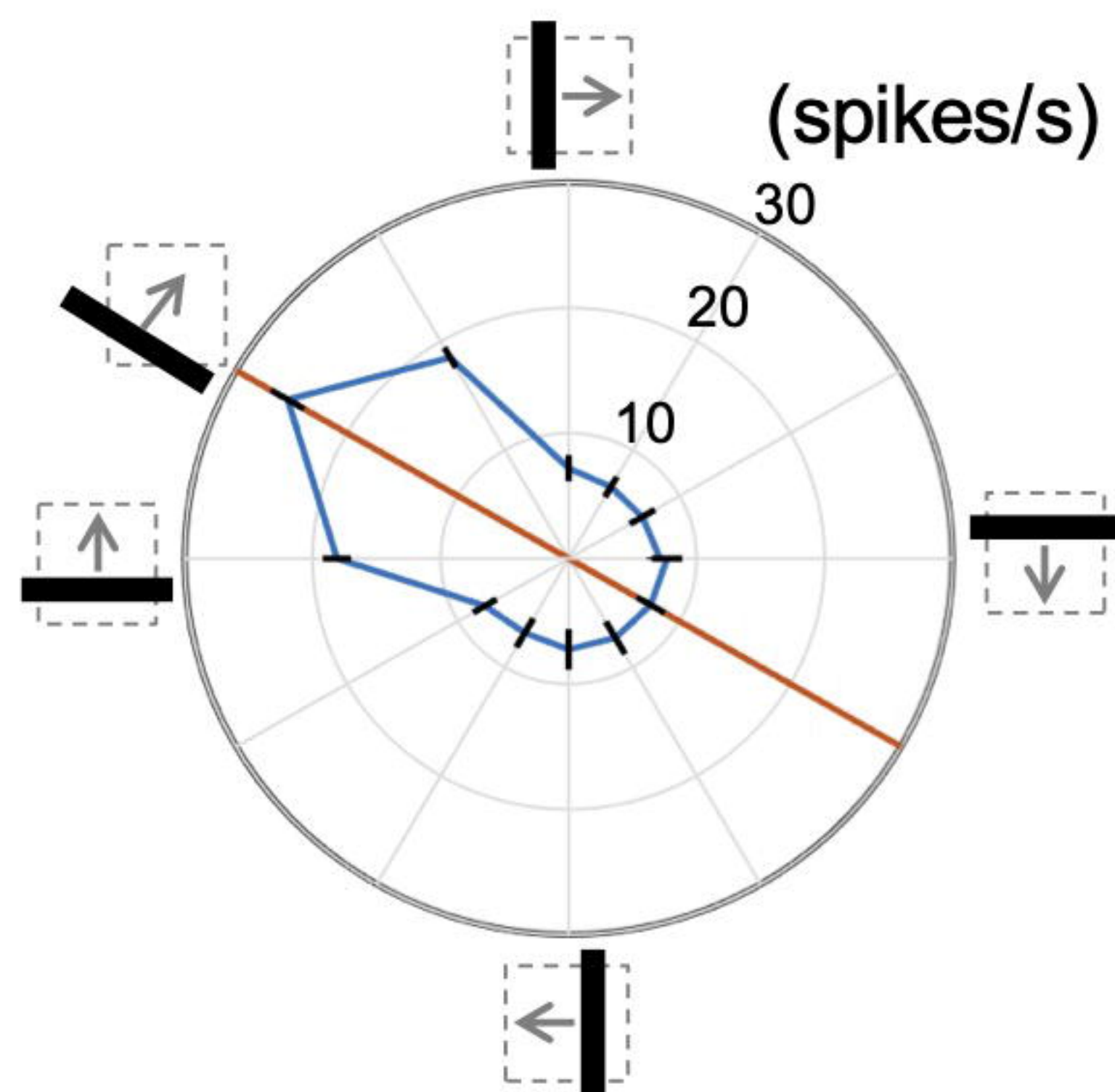
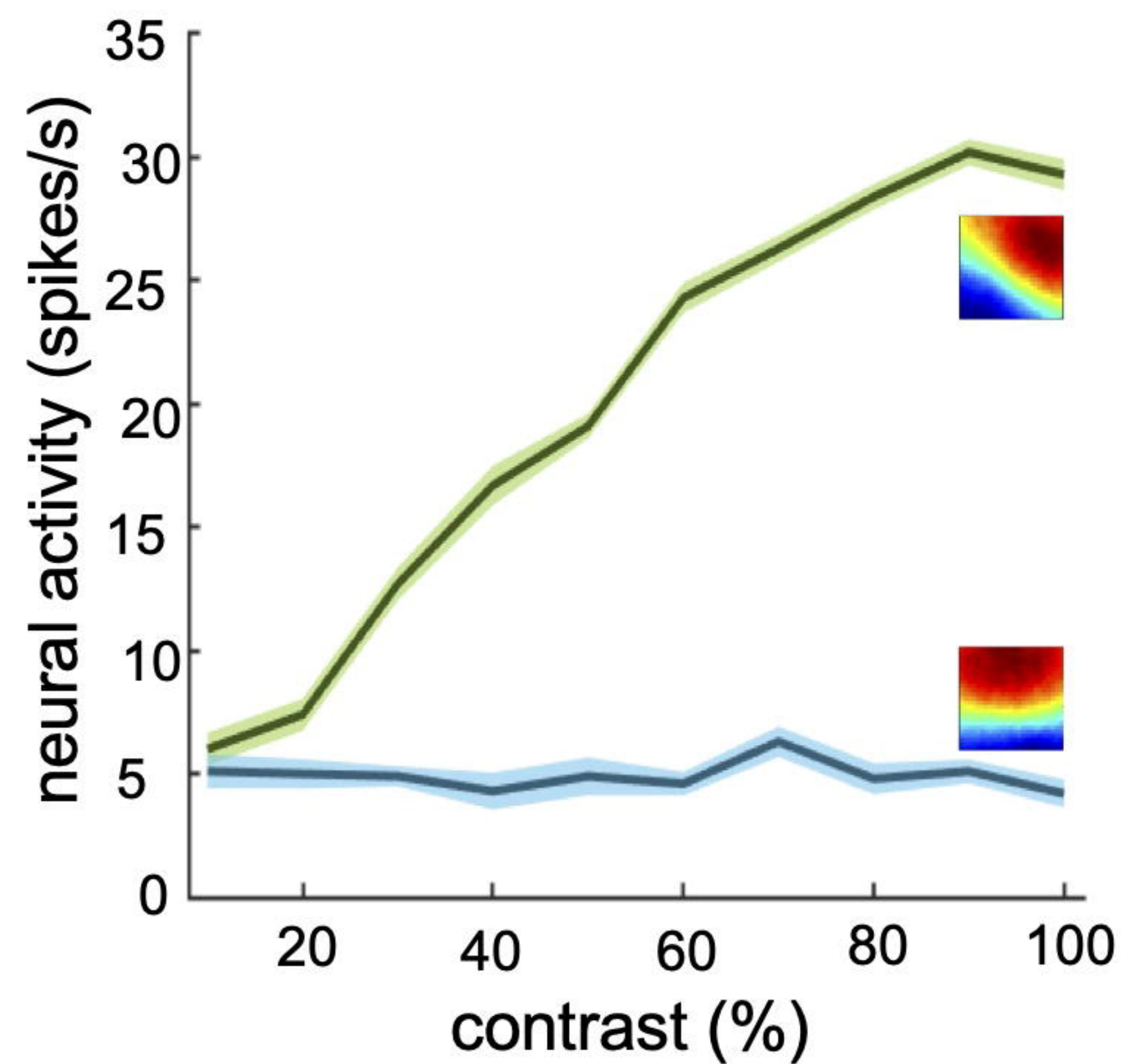
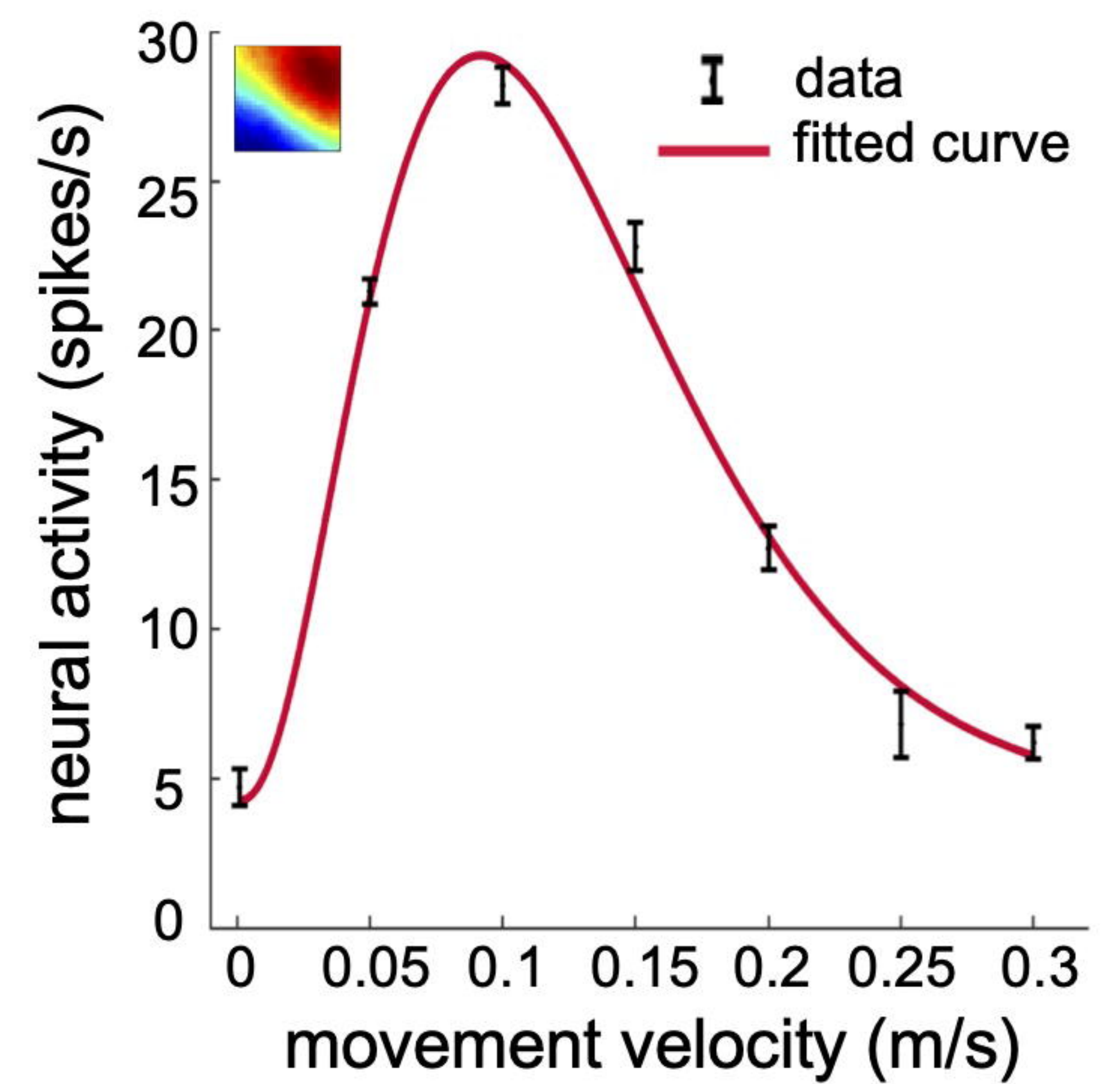








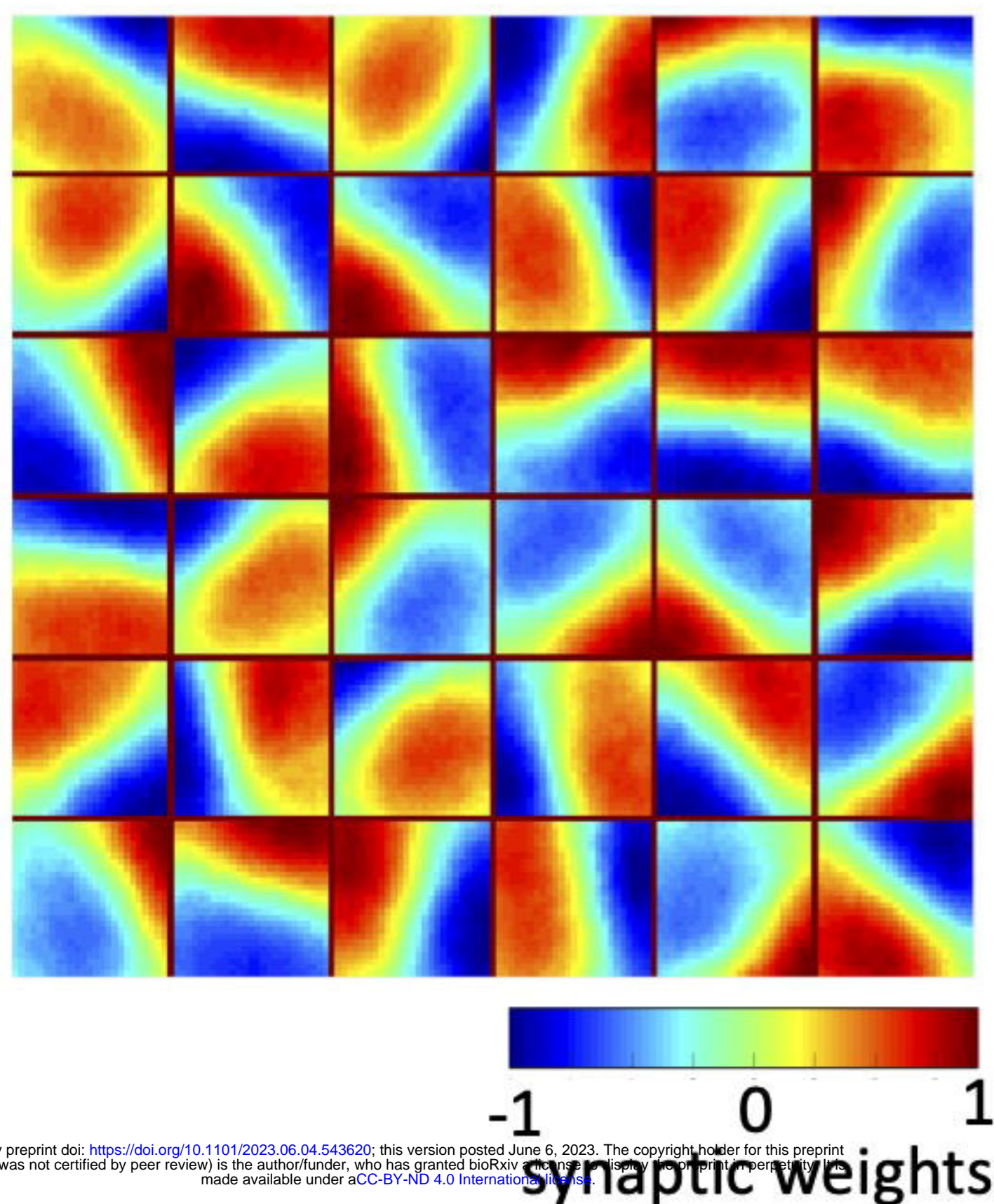


**A****B****C****D****E**

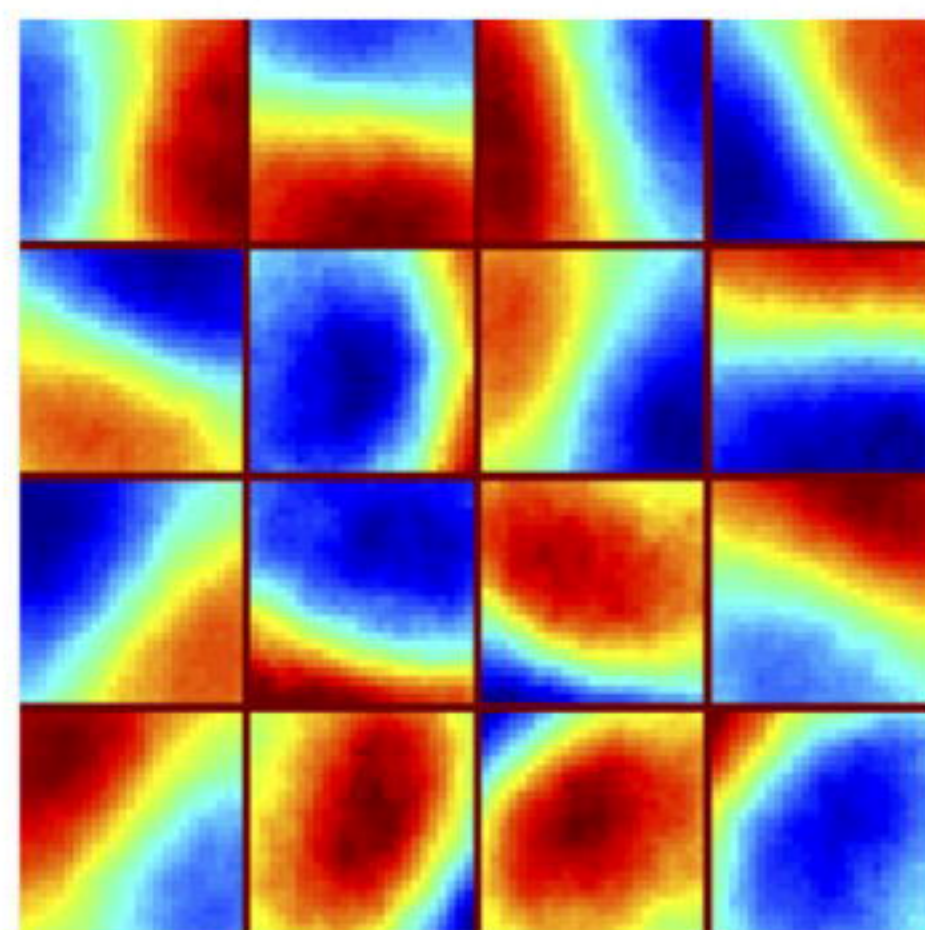


# A

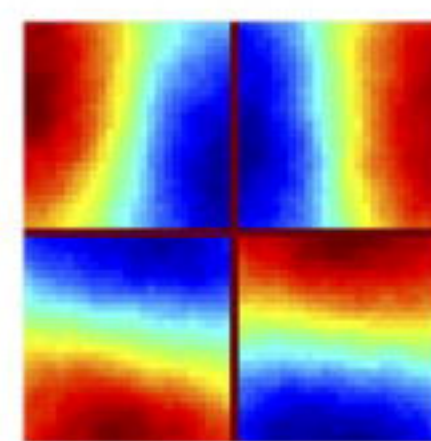
## model with 36 lobula neurons



## model with 16 lobula neurons

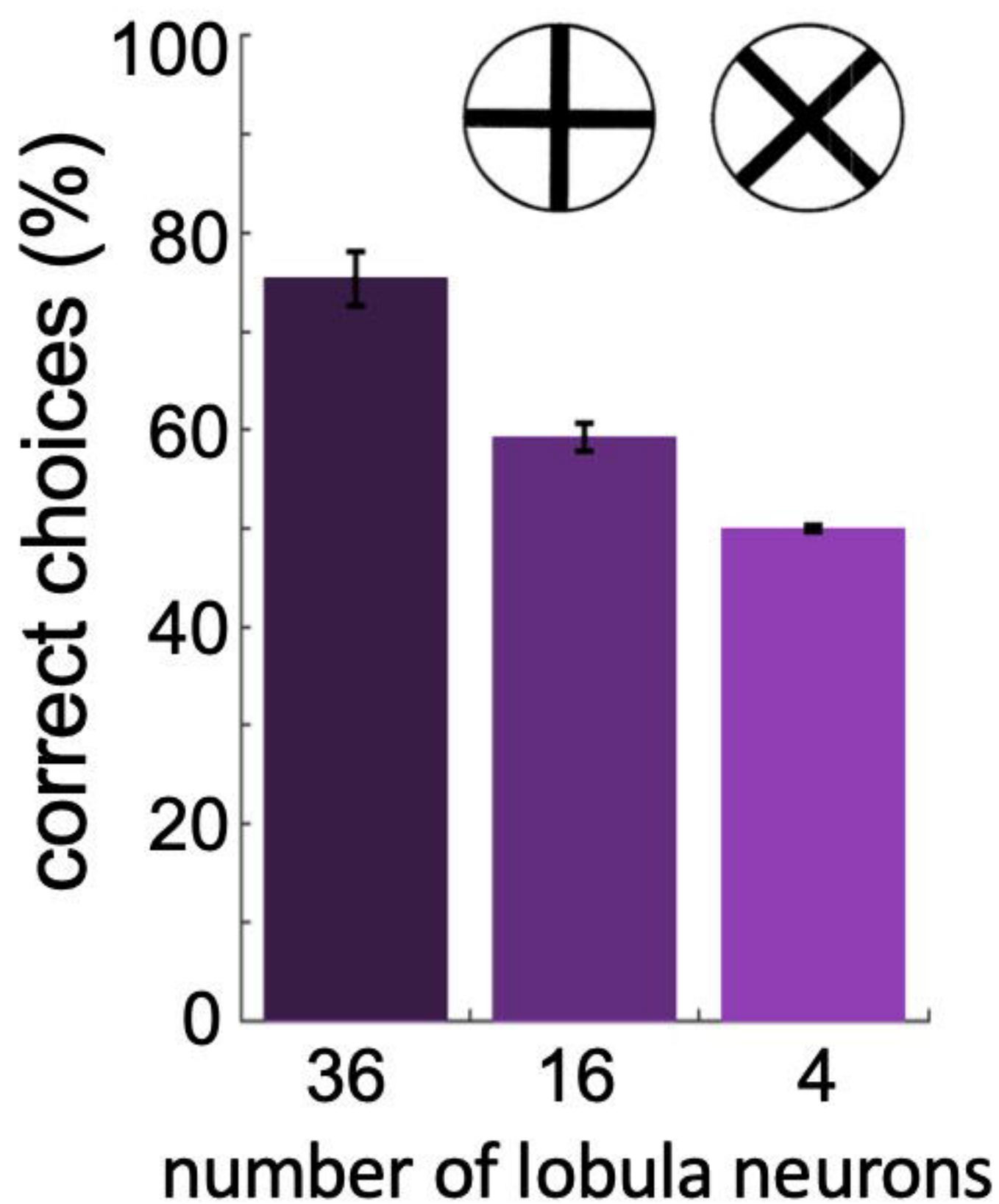


## model with 4 lobula neurons



bioRxiv preprint doi: <https://doi.org/10.1101/2023.06.04.543620>; this version posted June 6, 2023. The copyright holder for this preprint (which was not certified by peer review) is the author/funder, who has granted bioRxiv a license to display the preprint in perpetuity. It is made available under aCC-BY-ND 4.0 International license.

# B



# C

